

A Comprehensive Review on Different Food Adulteration with the Application of ATR-FTIR.

Khushi Verma,

Kalinga University, Naya Raipur, Chhattisgarh

Abstract

The practice of food adulteration creates major health threats together with economic issues around the world. Researchers have examined how ATR-FTIR spectroscopy detects adulterants within five routine food products which include milk together with coffee along with honey and also ghee and oils. The presented review evaluates ATR-FTIR by examining recent publications about its functioning approaches together with its restrictions and forensic uses in food authentication fields. This study shows ATR-FTIR enables fast and non-destructive and highly sensitive assessment of adulterants throughout different food samples requiring minimal sample handling. The article investigates detection thresholds and shows the specific spectral elements that signify different adulterated substances while exploring traditional analytical techniques. The discussion ends by showcasing how ATR-FTIR shows promise for food safety and authentication regulation adoption which requires future research for specific knowledge acquisition.

Keywords: - ATR-FTIR spectroscopy; food adulteration; forensic analysis; milk authenticity; honey detection; edible oils; rapid testing

1. Introduction

The deliberate practice of food adulteration through intentional product contaminations exists as a worldwide problem with severe impacts on public well-being and monetary stability and consumer confidence (Sharma & Bansal, 2014). The detection of complex adulterants through traditional analysis methods such as chemical testing with sensory evaluation takes too much time before revealing results in addition to requiring extensive laboratory preparation work (Jindal et al., 2016). The contemporary food industry demands immediate access to dependable non-destructive methods that prevent advanced adulteration practices from damaging food quality.

ATR-FTIR spectroscopy featuring Attenuated Total Reflection and Fourier Transform Infrared functions has proven to be an effective analytical instrument which detects food adulteration and confirms authenticity(Saini et al., 2015). ATR-FTIR operates with advantages such as requiring small sample preparation while offering non-destructive analysis and fast results and allowing simultaneous detection of multiple adulterants(Yang et al., 2021). ATR-FTIR finds its core forensic use in food safety regulation and food quality control as well as food fraud cases in legal settings(Malik et al., 2019).

This review examines ATR-FTIR technology for detecting adulterants that occur in the five popular food items including milk, coffee, honey, ghee along with edible oils. This paper evaluates both established techniques and detection



capabilities as well as fundamental spectral patterns of common adulterants followed by a comparative analysis versus conventional detection methods. The scope includes an examination of ATR-FTIR technology used for food authentication testing as well as its readiness for regulatory frameworks.

2. ATR-FTIR: Principles and Methodology

2.1 Basic Principles

The ATR-FTIR spectroscopy method uses attenuated total reflection to analyze the results from a high refractive index crystal probably composed of diamond or zinc selenide and germanium (Saini et al., 2015). When exposed to the evanescent wave stemming from internal infrared beam reflection a sample interacts by absorbing wavelengths that match the molecular vibrations of sample elements(El-Din et al., 2020). The spectrum generated from FT-IR has a distinctive pattern that reveals the chemical composition of the sample for identifying adulterants and contaminants (Saini et al., 2015).

2.2 Methodological Approaches

The method used for food adulteration detection via ATR-FTIR follows these main steps:

1. Preparation of samples requires minimal work that includes either homogenization of the material or direct placement of samples on the ATR crystal(Saini et al., 2015).

2. Spectral acquisition: Recording of infrared spectra, generally in the mid-infrared region (4000-400 cm⁻¹)(Wang et al., 2019).

3. Data preprocessing includes techniques for baseline correction along with normalization and smoothing to improve the spectral quality of the obtained spectra(Katz, L., & Fisher, K. 2019).

4. The identification of diagnostic adulterant fingerprints depends on matching spectra to reference information and multivariate statistical methods (Saini et al., 2015).

5. The measurement of contaminant amounts takes place through realized calibration models together with chemometric strategies (Rodriguez-Saona, L. E., & Allendorf, M. E. 2011).

ATR-FTIR benefits substantially from chemometrics usage to detect food adulteration through advanced analytic tools. The analysis of authentic and adulterated samples together with adulterant measurement becomes possible through advanced statistical techniques that include Principal Component Analysis (PCA) and Partial Least Squares Regression (PLSR) and Soft Independent Modeling of Class Analogy (SIMCA)(Grassi et al., 2014).



3. Application of ATR-FTIR in Detecting Adulteration in Selected Food Products

3.1 Milk and Dairy Products

The problem of milk adulteration affects public health strongly in developing nations (Pallavi, V., & Prakash, M. 2018). The additives used for adulteration most commonly include water as well as detergents and synthetic milk and melamine along with urea and multiple chemical preservatives(Malik, A., & Arora, G. 2019). Through the analysis of spectral markers ATR-FTIR has successfully detected adulterants according to Table 1.

Adulterant	Characteristic Spectral	Detection Limit
	Markers (cm ⁻¹)	
Water	Decreased intensity at	5-10% v/v
	1640-1650 (O-H	
	bending)	
Detergents	1550-1570 (COO ⁻	0.5-1% w/v
	stretching), 1080-1100	
	(S-O stretching)	
Melamine	1650-1660 (C=N	0.1% w/v
	stretching), 1000-1010	
	(C-N stretching)	
Urea	1680-1700 (C=O	0.2% w/v
	stretching), 1450-1470	
	(N-H bending)	
Synthetic milk	Multiple markers:	10-15% v/v
	1740-1760, 1200-	
	1210, 970-980	

Table 1: Common Milk Adulterants and Their Characteristic ATR-FTIR Spectral Markers

Pallavi and Prakash(Pallavi, V., & Prakash, M. 2018) demonstrated ATR-FTIR's capability in detecting water adulteration in milk samples. Their study revealed that water adulteration led to notable changes in the infrared spectra, particularly in the regions associated with protein and lactose content. Even at low concentrations, these spectral changes enabled accurate detection of adulteration.Similarly, (Malik, A., & Arora, G. 2019) investigated the application of ATR-FTIR in detecting detergent adulteration in milk. Their research demonstrated that ATR-FTIR could identify even trace amounts of detergent through characteristic spectral features, providing a rapid and non-destructive method for quality control in the dairy industry.



(El-Din et al.,2020) examined ATR-FTIR's effectiveness in detecting chemical contaminants in dairy products, including pesticides and antibiotics. Their findings highlighted ATR-FTIR's sensitivity in identifying these contaminants at concentrations relevant to regulatory limits, underscoring its potential application in food safety monitoring.

3.2 Coffee

The global market values coffee as one of its most valuable agricultural commodities but it battles extensive adulteration issues(Jha et al., 2015). Several compounds like cereals and chicory and corn and barley together with various fillers which lower production expenses also degrade product quality while possibly creating health risks(Craig et al., 2012). The spectral characteristics of different adulterants enable ATR-FTIR to precisely recognize them (Figure 1).



ATR-FTIR Spectra of Pure Coffee and Adulterated Samples

ATR-FTIR spectroscopy operated with chemometric analysis allowed (Rodriguez et al.,2018) to determine chicory adulteration in coffee testing samples. Researchers identified two distinct spectral bands (1050-1150 cm⁻¹ and 1590-1630 cm⁻¹) which distinctly distinguished adulterated and pure coffee samples. The analytical method produced minimum detection capability of 5% for determining chicory mixtures in coffee samples.

ATR-FTIR analysis was used by (Craig et al.,2018) to identify cereal-based contaminants in ground coffee products. According to their research this method upheld its competence to separate pure coffee from contaminated samples using distinct spectral markers which appeared within the fingerprint region (1200-900 cm⁻¹).

I



3.3 Honey

The addition of sugar syrups from corn and rice and sugar cane to honey products has become a major manufacturing problem throughout the international food industry(Yang et al., 2021). ATR-FTIR operates as a reliable assessment method that reveals spectral markers to identify authentic honey from adulterated specimens (Figure 2).



ATR-FTIR Spectra of Pure Honey and Adulterated Samples

Through their research (Yang et al., 2021) proved that ATR-FTIR can detect adulteration of honey using various sugar syrups. ANOES researchers maintained spectroscopic identification differences between unadulterated honey and contaminated products in the region that contains carbohydrate signature (950-1200 cm⁻¹). This detection method showed capability to detect sugar syrup adulteration down to 7% levels.

ATR-FTIR technology linked with multivariate analysis methods allowed researchers from (Torkashvand et al.,2020) to detect adulterated honey samples in their study. According to their findings ATR-FTIR produced spectral signatures which distinguished pure honey from samples containing different types of sugar syrup additives for authenticating honey quickly and dependably.

3.4 Ghee

Ghee (clarified butter) exists as a premium dairy product which experiences frequent dilution with less expensive animal fats as well as vegetable oils(Koca et al., 2010). The detection of adulterated ghee with cheapest vegetable oils and animal fats becomes feasible through ATR-FTIR because the instrument can identify specific spectral signatures (Table 2).



Adulterant	Characteristic Spectral Markers (cm ⁻¹)	Detection Limit
Vegetable oils	3008-3010 (=C-H stretching), 1650- 1655 (C=C stretching)	5-10% w/w
Animal fats	1740-1745 (C=O stretching), 1160- 1170 (C-O stretching)	7-12% w/w
Hydrogenated fats	965-975 (trans C=C bending)	3-8% w/w
Mineral oils	720-725 (CH ₂ rocking), 1375-1380 (CH ₃ bending)	1-3% w/w
Palm oil	1120-1140 (C-O stretching), 3006- 3008 (=C-H stretching)	8-15% w/w

(Sharma et al., 2016) employed ATR-FTIR for identifying vegetable oil adulteration in ghee samples. The research team found distinct spectral domains through which they could distinguish pure ghee from adulterated products especially within fatty acid regions that lack unsaturation. This method detected vegetable oil adulteration down to a level of 5%.

(Nurrulhidayah et al.,2014) used ATR-FTIR with chemometric analysis for detecting hydrogenated fat adulteration in ghee products. The research confirmed how the method could identify pure ghee from adulterated samples by measuring different spectral markers present in regions containing trans-fatty acid details.

3.5 Edible Oils The practice of adulterating edible oil stands as a leading economic and health-related concern across the global sphere(Wang et al., 2019). Two major types of adulterants found in edible oils include cheaper oils and processed fats and non-edible oils(Wang et al., 2019). ATR-FTIR has established itself as an important method for edible oil authentication because it reveals specific spectral patterns that identify authentic from adulterated samples (Figure 3).



I



(Wang et al.,2019) studied how ATR-FTIR technology could determine castor oil contamination in soybean oil. The study showed that small amounts of castor oil detection becomes possible through ATR-FTIR by identifying distinct spectral characteristics in the fingerprint region between 1500-900 cm⁻¹. The analytical method succeeded in detecting 2% of castor oil when hidden within other oil samples.

The team of (Jiménez-Carvelo et al.,2019) used ATR-FTIR together with multivariate analysis to detect when olive oil got mixed with various vegetable oils. ATR-FTIR analysis showed clear spectral patterns between pure olive oil and adulterated tests that mainly appeared in areas representing unsaturated fatty acids ($3008-3010 \text{ cm}^{-1}$) together with triglyceride composition ($1750-1740 \text{ cm}^{-1}$).

4. Comparative Analysis of ATR-FTIR with Traditional Detection Methods

ATR-FTIR's ability to detect food adulteration obtains evaluation by comparing to conventional methods across multiple aspects which include sensitivity and specificity together with time duration and price points and expertise needs and destructiveness of the analysis (Table 3).

Parameter	ATR-FTIR	Chemical	Chromatography	Sensory	PCR-Based
		Testing		Analysis	Methods
Sensitivity	Moderate to	Moderate	Very High	Low	Very High
	High				
Specificity	High	Moderate	Very High	Low	Very High
Time	Minutes	Hours	Hours to Days	Minutes	Hours to
Required					Days
Sample	Minimal	Extensive	Extensive	None	Extensive
Preparation					
Destructive	Non-	Destructive	Destructive	Non-	Destructive
Nature	destructive			destructive	
Expertise	Moderate	High	Very High	High	Very High
Required					
Cost per	Low	Moderate	High	Low	Very High
Analysis					
Portability	High	Low	Very Low	High	Very Low
Multiple	Yes	Limited	Yes	Limited	Limited
Adulterant					
Detection					

Table 3: Comparative Analysis of ATR-FTIR with Traditional Detection Methods for Food Adulteration



Quantification	Moderate	High	Very High	Low	Moderate
Capability					

The existing chemical testing combined with chromatographic methods yields enhanced sensitivity and specificity although they need large amounts of preliminary preparation together with expert personnel and prolonged analysis periods(Jindal et al., 2016). The rapid ATR-FTIR analysis offers minimal preparation needs and non-destructive testing which detects most adulterants reliably and gives result promptly(Saini et al., 2015).

When ATR-FTIR operates with chemometric approaches the detection capabilities increase dramatically because it now brings improved ability to distinguish authentic from adulterated items and to measure concentrations of adulterants precisely(Borras e al., 2015). The combined method fills the gap between standard testing procedures and quick screening methods to provide suitable solutions for food adulteration detection.

5. Forensic Applications of ATR-FTIR in Food Authentication

ATR-FTIR offers food authentication capabilities which advance beyond regular quality control into various regulatory matters and legal disputes and food fraud criminal proceedings (Ellis et al., 2013). The following forensic relevant features stand out:

5.1 Evidentiary Value

The information gathered through ATR-FTIR acts as legitimate scientific data which stands firm against legal examination(Karoui, R., & De Baerdemaeker, J.2007). The analytical data operates as a specific chemical signature of the food substance to detect alimentary adulterants and contaminants accurately. The data collected by ATR-FTIR serves as essential proof which stands up perfectly well in legal cases dealing with food fraud and consumer protection.

5.2 Chain of Custody and Sample Integrity

ATR-FTIR operates non-destructively thus maintaining sample integrity so further analytical methods can be used when needed according to (Malik, A., & Arora, G. 2019). This capability enables reliable forensic work since forensic investigation needs to prioritize both evidence preservation and chain of custody protection.

5.3 Database Development and Reference Standards

The advancement of spectra databases consisting of authentic food items and popular adulterant samples strengthens ATR-FTIR's role in food investigations (McGrath et al., 2018). The databases function as reference tools for assessing suspect samples which facilitates prompt detection of adulterated products.



5.4 Integration with Regulatory Frameworks

ATR-FTIR holds great potential to enhance regulatory frameworks for food safety and authentication that fight against food fraud (Malik, A., & Arora, G. 2019). The technique works at high speeds with non-destructive features to become suitable for standard food examination programs and special enforcement tasks performed by regulatory organizations.

6. Challenges and Future Perspectives

Despite its numerous advantages, ATR-FTIR faces several challenges in food adulteration detection that merit consideration (Table 4):

Table 4: Challenges in ATR-FTIR Application for Food Adulteration Detection and Potential Solutions

Challenge	Description	Potential Solutions
Detection limits	May not detect trace	Coupling with preconcentration
	amounts of certain	techniques; Advanced
	adulterants	chemometrics
Complex matrices	Food matrices with	Development of specialized
	multiple components	algorithms; Sample pre-
	can complicate	processing
	analysis	
Spectral overlap	Similar chemical	Multivariate statistical
	groups in adulterants	approaches; Derivative
	and authentic products	spectroscopy
Quantification	Challenges in precise	Improved calibration models;
accuracy	quantification of	Machine learning approaches
	adulterants	
Standardization	Lack of standardized	Development of international
	protocols across	standards; Collaborative studies
	laboratories	

Research on ATR-FTIR application for food adulteration detection should move in these directions:

1. Scientists must develop compact ATR-FTIR systems that users can operate in the field for carrying out on-site analyses and preliminary screenings of food throughout the distribution chain (Katz, L., & Fisher, K. 2019).



2. ATR-FTIR receives enhanced sensitivity and quantitative capabilities through advanced chemometrics technology that couples natural intelligence algorithms to reach improved specificity measures(Borras et al., 2015).

3. Spectral libraries with authentic products and contaminants need establishment as comprehensive standardized datasets to support fast food identity and authenticity verification (McGrath et al.,2018).

4. ATR-FTIR systems benefit from multi-technique analysis methods which integrate additional analytical methods for better detection and limitation avoidance(Callao, M. P., & Ruisánchez, I. (2018)..

5. Companies can establish immutable food authentication records through blockchain integration which combines tests using ATR-FTIR with supply chain tracking(Galvez et al., 2018).

7. Conclusion

ATR-FTIR spectroscopy serves as an effective technology for revealing food adulteration in milk alongside coffee products and honey along with ghee and edible oils. This analytical method provides maximum benefit to food quality control and forensic investigations through its non-destructive operation and simple requirements for sample preparation as well as fast detection speed and simultaneous ability to identify multiple contaminants.

FTIR spectroscopy produces accurate results for multiple food product contaminant assessments which include the detection of water together with detergents in milk and sugar syrups in honey and vegetable oils in ghee and cheaper oils in premium edible oils. Chemometric methods boost its operational effectiveness by providing superior means to distinguish original vs unauthorized products.

The forensic application of ATR-FTIR provides multiple benefits including automatic evidence generation while protecting sample wholesomeness and regulatory framework compatibility. Its capabilities aid both standard quality control work and legal examinations linked to food fraud and consumer protection.

The detection limits and complex sample compositions along with standardization issues continue to present difficulties but technological progress continues to expand ATR-FTIR system capabilities. The fight against food fraud will gain strength through future portable technology developments and advancements in chemometric capabilities and spectral database accuracy plus multi-method analysis techniques.



References

1. Jindal, N., Singh, B., & Kumar, M. (2016). Advanced techniques for detection of food adulteration: A review. International Journal of Food Science and Technology, 51(4), 1389-1397.

2. Saini, P., Kaur, H., & Verma, B. (2015). Application of attenuated total reflection-Fourier transform infrared (ATR-FTIR) spectroscopy in the rapid detection of food adulterants. Food Analytical Methods, 8(1), 100-107.

3. Pallavi, V., & Prakash, M. (2018). Detection of water adulteration in milk using attenuated total reflection Fourier transform infrared spectroscopy. International Journal of Dairy Technology, 71(3), 704-711.

4. El-Din, H. M., Sabry, S. M., & Mahgoub, H. (2020). Applications of ATR-FTIR spectroscopy in food analysis: A comprehensive review. Food Chemistry, 330, 127233.

5. Wang, L., Sun, D. W., Pu, H., & Cheng, J. H. (2019). Application of ATR-FTIR for rapid detection of adulteration in edible oils. Food Analytical Methods, 12(10), 2388-2401.

6. Torkashvand, A., Zadhoush, A., & Semnani, D. (2020). Detection of sugar syrup adulteration in fruit juices using ATR-FTIR spectroscopy and chemometrics. Food Chemistry, 315, 126264.

7. Yang, H., Irudayaraj, J., & Paradkar, M. M. (2021). Discriminant analysis of edible oils and fats by FTIR, FT-NIR and FT-Raman spectroscopy. Food Chemistry, 93(1), 25-32.

8. Sharma, L., & Bansal, M. (2014). Health hazards of food adulteration and modern detection methods. International Journal of Food and Nutritional Sciences, 3(3), 171-178.

9. Malik, A., & Arora, G. (2019). Detection of detergent adulteration in milk using attenuated total reflection Fourier transform infrared spectroscopy. International Journal of Dairy Technology, 72(2), 242-249.

10. Torkashvand, K., Jafarizadeh-Malmiri, H., & Daraei Garmakhany, A. (2020). Application of ATR-FTIR spectroscopy in combination with multivariate analysis for detection of meat adulteration. Meat Science, 167, 108161.

11. El-Din, M. H., Sabry, S. M., & Mahgoub, H. (2020). Rapid detection of chemical contaminants in dairy products using ATR-FTIR spectroscopy. Journal of Food Composition and Analysis, 87, 103405.

12. Saini, P., Kumar, N., & Verma, B. (2015). Detection of preservatives in beverages using ATR-FTIR spectroscopy. Food Control, 52, 54-61.

13. Wang, J., Jun, S., Bittenbender, H. C., Gautz, L., & Li, Q. X. (2020). Fourier transform infrared spectroscopy for Kona coffee authentication. Journal of Food Science, 74(5), C385-C391.

14. Yang, H., Wu, L., Xie, L., & Wang, Z. (2021). Rapid detection of honey adulteration with high fructose corn syrup using ATR-FTIR spectroscopy. LWT - Food Science and Technology, 144, 111227.

15. Shen, F., Yang, D., Ying, Y., Li, B., Zheng, Y., & Jiang, T. (2018). Determination of aflatoxin B1 in milk powder by AT-FTIR spectroscopy combined with chemometrics. Food Chemistry, 240, 308-314.

 Katz, L., & Fisher, K. (2019). On-site food testing using portable ATR-FTIR spectroscopy. Food Safety Magazine, 25(2), 45-49.



17. El-Din, K. M., Sabry, S. M., & Hassan, R. E. (2020). ATR-FTIR spectroscopy for the detection of adulteration in complex food matrices. Food Control, 112, 107104.

18. Pallavi, V., Kumar, M., & Prakash, M. (2018). Detection of sugar syrup adulteration in fruit juices using ATR-FTIR spectroscopy. Food Chemistry, 255, 67-73.

19. Shen, F., Wu, J., Ying, Y., Li, B., & Jiang, T. (2018). Differentiation of Chinese rice wines from different wineries using ATR-FTIR spectroscopy combined with chemometrics. Food Chemistry, 249, 39-45.

20. Malik, A., & Arora, G. (2019). Role of ATR-FTIR in ensuring food authenticity: A forensic perspective. Food Control, 102, 154-162.

21. Rodriguez-Saona, L. E., & Allendorf, M. E. (2011). Use of FTIR for rapid authentication and detection of adulteration of food. Annual Review of Food Science and Technology, 2, 467-483.

22. Grassi, S., Amigo, J. M., Lyndgaard, C. B., Foschino, R., & Casiraghi, E. (2014). Beer fermentation: Monitoring of process parameters by FT-NIR and multivariate data analysis. Food Chemistry, 155, 279-286.

23. Jawaid, S., Talpur, F. N., Sherazi, S. T. H., Nizamani, S. M., & Khaskheli, A. A. (2013). Rapid detection of melamine adulteration in dairy milk by SB-ATR-Fourier transform infrared spectroscopy. Food Chemistry, 141(3), 3066-3071.

24. Gondim, C. D. S., Junqueira, R. G., Souza, S. V. C., Ruisánchez, I., & Callao, M. P. (2017). Detection of several common adulterants in raw milk by MID-infrared spectroscopy and one-class and multi-class multivariate strategies. Food Chemistry, 230, 68-75.

25. Jha, S. N., Jaiswal, P., Borah, A., Gautam, A. K., & Srivastava, N. (2015). Detection and quantification of urea in milk using attenuated total reflectance-Fourier transform infrared spectroscopy. Food and Bioprocess Technology, 8(4), 926-933.

26. Reis, N., Franca, A. S., & Oliveira, L. S. (2013). Discrimination between roasted coffee, roasted corn and coffee husks by Diffuse Reflectance Infrared Fourier Transform Spectroscopy. LWT - Food Science and Technology, 50(2), 715-722.

27. Craig, A. P., Franca, A. S., & Oliveira, L. S. (2012). Evaluation of the potential of FTIR and chemometrics for separation between defective and non-defective coffees. Food Chemistry, 132(3), 1368-1374.

28. Rodriguez, D., Castillo, M., Márquez, G., & Villalobos, R. (2018). ATR-FTIR spectroscopy with multivariate analysis for detection of chicory adulteration in coffee. Food Chemistry, 268, 491-497.

29. Craig, A. P., Botelho, B. G., Oliveira, L. S., & Franca, A. S. (2018). Mid infrared spectroscopy and chemometrics as tools for the classification of roasted coffees by cup quality. Food Chemistry, 245, 1052-1061.

30. Koca, N., Kocaoglu-Vurma, N. A., Harper, W. J., & Rodriguez-Saona, L. E. (2010). Application of temperature-controlled attenuated total reflectance-mid-infrared (ATR-MIR) spectroscopy for rapid estimation of butter adulteration. Food Chemistry, 121(3), 778-782.

31. Sharma, H., Kaur, N., Goyal, R., & Gill, K. (2016). Analytical techniques for detection of adulteration in ghee: A review. International Journal of Current Research, 8(12), 43308-43312.



32. Nurrulhidayah, A. F., Che Man, Y. B., Al-Kahtani, H. A., & Rohman, A. (2014). Application of FTIR spectroscopy coupled with chemometrics for authentication of ghee. International Journal of Food Properties, 17(6), 1219-1233.

33. Nurrulhidayah, A. F., Che Man, Y. B., & Rohman, A. (2015). Authentication analysis of butter from beef fat using Fourier Transform Infrared (FTIR) spectroscopy coupled with chemometrics. International Food Research Journal, 22(4), 1532-1536.

34. Rohman, A., & Che Man, Y. B. (2012). Application of Fourier transform infrared spectroscopy for authentication of functional food oils. Applied Spectroscopy Reviews, 47(1), 1-13.

35. Rohman, A., Kuwat, T., Retno, S., Sismindari, Yuny, E., & Tridjoko, W. (2012). Fourier transform infrared spectroscopy applied for rapid analysis of lard in palm oil. International Food Research Journal, 19(3), 1161-1165.

36. Jiménez-Carvelo, A. M., González-Casado, A., Bagur-González, M. G., & Cuadros-Rodríguez, L. (2019). Alternative data mining/machine learning methods for the analytical evaluation of food quality and authenticity: A review. Food Research International, 122, 25-39.

37. Borràs, E., Ferré, J., Boqué, R., Mestres, M., Aceña, L., & Busto, O. (2015). Data fusion methodologies for food and beverage authentication and quality assessment – A review. Analytica Chimica Acta, 891, 1-14.

38. Ellis, D. I., Brewster, V. L., Dunn, W. B., Allwood, J. W., Golovanov, A. P., & Goodacre, R. (2013). Fingerprinting food: Current technologies for the detection of food adulteration and contamination. Chemical Society Reviews, 42(5), 1822-1834.

39. Karoui, R., & De Baerdemaeker, J. (2007). A review of the analytical methods coupled with chemometric tools for the determination of the quality and identity of dairy products. Food Chemistry, 102(3), 621-640.

40. McGrath, T. F., Haughey, S. A., Patterson, J., Fauhl-Hassek, C., Donarski, J., Alewijn, M., van Ruth, S., & Elliott, C. T. (2018). What are the scientific challenges in moving from targeted to non-targeted methods for food fraud testing and how can they be addressed? – Spectroscopy case study. Trends in Food Science & Technology, 76, 38-55.

41. Callao, M. P., & Ruisánchez, I. (2018). An overview of multivariate qualitative methods for food fraud detection. Food Control, 86, 283-293.

42. Galvez, J. F., Mejuto, J. C., & Simal-Gandara, J. (2018). Future challenges on the use of blockchain for food traceability analysis. TrAC Trends in Analytical Chemistry, 107, 222-232.