

# Assessment of Fluctuation in Aeromycological Diversity at Different Sites in Gorakhpur District, Uttar Pradesh, India

Ujjawal Prakash<sup>1</sup>, Rajneesh Rao<sup>1</sup>, Sanyogita Kumari<sup>2</sup> and Ashok Kumar<sup>1\*</sup>

<sup>1</sup>Department of Botany, Deen Dayal Upadhyaya Gorakhpur University,  
Gorakhpur-273009, India

<sup>2</sup>Department of Botany, Raghunath Girl's Post Graduate College, Meerut-250001, India

## Abstract

The purpose of the current study's aeromycological survey was to determine the effect of temperature and relative humidity within a day on prevalence of aeromycoflora diversity in different locations in Gorakhpur district, during first week of October in 2023. For sampling, the gravity-based Petri-plate approach using Potato Dextrose Agar (PDA) medium was employed to trap fungal spores and mycelial fragments. During study a total 21 identified species belonging to 13 fungal genera were isolated excluding Mucorales, mycelia sterilia and unidentified fungi. The findings reveal that a total 1506 and 2287 fungal isolates were recovered from air sampling of study sites during morning and late afternoon respectively. The highest mycoflora occurrence frequency (%) was found in Kushmi forest both in morning (19.46%) and late afternoon (22.60%) while, lowest in morning (6.30%) and late afternoon (7.08%) was determined in Baba Raghav Das (BRD) medical college and AIIMS (All India Institute of Medical Sciences), Gorakhpur respectively. Collectively, during morning sampling, *Cladosporium* sp. exhibited highest relative density (19.85%) followed by *Aspergillus flavus* (17.13%), *Aspergillus niger* (16.73%) and *Penicillium* sp. (10.89%), whereas, both *Acremonium* sp. (0.13%) and *Colletotrichum* sp. (0.13%) having lowest relative density followed by *Alternaria atra* (0.20%), and *Alternaria longipes* (0.33%). The late afternoon sampling reveals, *Cladosporium* sp. had highest relative density (22.39%) followed by *Aspergillus niger* (17.13%), *Aspergillus flavus* (16.73%) and *Penicillium* sp. (10.89%). Among all fungal spore types, genus *Aspergillus* showed dominancy in total spore count i.e. 700 (46.48%) and 1134 (49.58%) both in morning and late afternoon respectively. The spore count in air from morning to late afternoon enhanced with increase of temperature in major mycoflora viz. *Cladosporium* sp. (299 to 512), *Aspergillus niger* (252 to 491), *Aspergillus flavus* (258 to 349) and *Penicillium* sp. (164 to 185), strengthen the effect of meteorological factors (temperature, relative humidity etc) along with anthropogenic factors on aeromycoflora diversity.

**Keywords:** Aeromycobiota; Meteorological Factors; Library; Forest; Temple; Fungal isolates

## 1. Introduction

Air is crucial component of the environment that contains many abiotic and biotic entities, mandatory for existence of life. The biotic components of air, collectively known as 'bioaerosol' are minute biological particles suspended in atmosphere playing crucial role in various ecological, environmental and health related processes (Estill et al., 2016; Kim et al., 2018). All the major groups of microbes including viruses, bacteria, fungi etc. are present in the atmosphere (Tastassa et al., 2024). Aeromycoflora includes fungal spores and microscopic mycelial fragments, which can be dispersed through the air over long distances (Adams et al., 2013). Fungal spores constitute a major component of aerobiota (Niu et al., 2021). The sources of mycoflora in air may be natural e.g. soil, decaying organic matter, water bodies and vegetation or anthropogenic e.g. agricultural activities, waste disposal etc. (Prussin & Marr, 2015). The air temperature, precipitation, relative humidity, geographic distribution, atmospheric turbulence, wind speed,

quantity of organic matter and UVB radiation are some of the meteorological factors that affect the sporulation and dispersal of fungi (Crandall & Gilbert, 2017; Grinn-Gofroń et al., 2018; van Rhijn et al., 2021; Ajikah et al., 2023).

Airborne fungi also play a vital role in ecological balance, agriculture, and various industries. They significantly contribute to nutrient cycling by decomposing organic matter to enrich soil fertility, and supporting plant growth through mycorrhizal associations (Park et al., 2024). Many airborne fungi are essential in industrial biotechnology to produce antibiotics, enzymes, organic acids and also used in pharmaceuticals and food processing. Additionally, few aeromycobiota play a key role in biocontrol, to suppress plant pathogens by potent antagonistic activity (Madsen et al., 2007). Certain aeromycoflora are valuable in bioremediation, breaking down pollutants and toxic substances, thereby helping to clean the air and soil (Vaksmas et al., 2023). Their ability to spread through the air ensures their widespread benefits in maintaining biodiversity but on the other hand certain airborne fungal spores can also cause respiratory allergies, infections, and crop diseases, impacting agriculture and public health.

Airborne fungi are also key indicators of atmospheric bio-pollution level (Pusz et al., 2018; Castro e Silva et al., 2020), but, their absolute consequences are still unclear. Breathing in bioaerosol having airborne microbes and their by-products, can cause respiratory diseases along with other health issues such hypersensitivity, infections, pneumonia and toxic responses (Kim et al., 2018; Durugbo et al., 2023). Currently, over 80 fungal genera have been recognized as aeroallergens linked with respiratory tract (Rick et al., 2016; Letovsky et al., 2024). Species of many aerofungi, including *Aspergillus*, *Alternaria*, *Cladosporium*, *Fusarium*, *Penicillium* etc. may induce allergic responses, sore throat, severe asthma, fatigue, headache and irritation of eye and sinus in human (Oliveira et al., 2023).

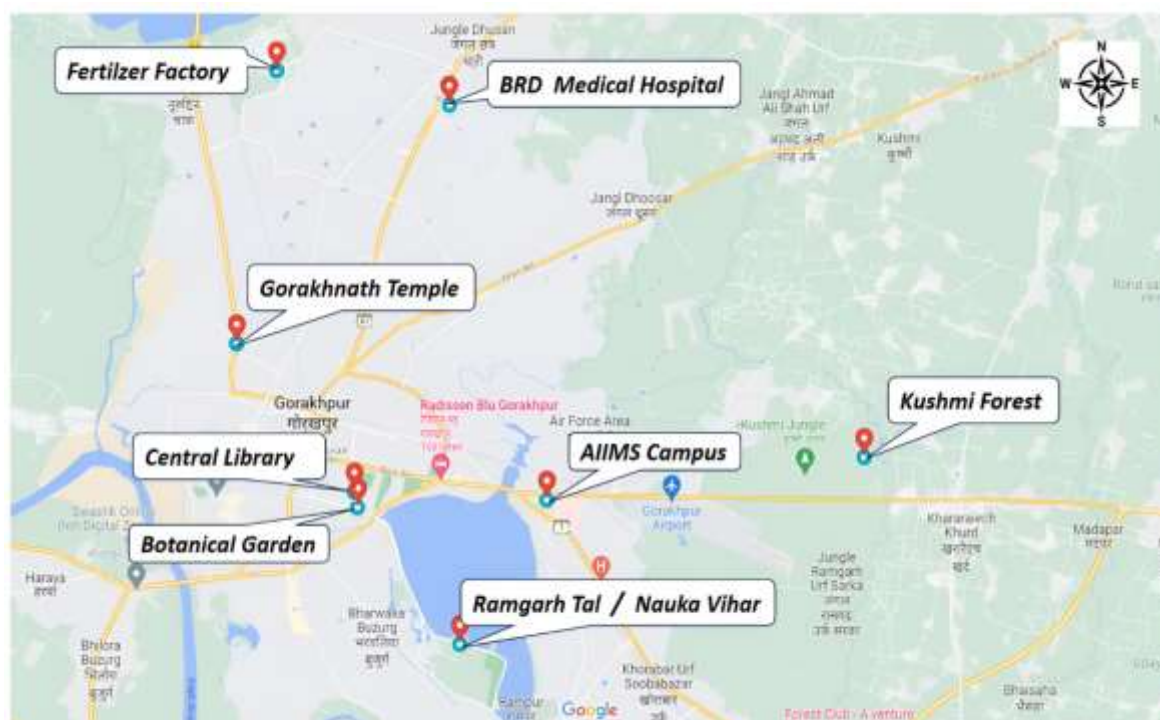
Several aeromycological studies in India are crucial due to the country's diverse climate, vegetation, environmental conditions, and geographic area which influence airborne fungal spore distribution (Vijayalakshmi et al., 2020). Seasonal changes also have great impact on it (Kochar et al., 2014). Research across various regions has identified allergenic and pathogenic fungi affecting public health, agriculture, and air quality (Pavan & Manjunath, 2014; Baxi et al., 2016). Studies in urban and rural areas reveal seasonal variations in fungal spores, with higher concentrations during monsoon and post-harvest periods (Kakde et al., 2001).

Aeromycological studies offer novel understanding about the diversity, distribution, and dynamics of airborne fungal spores, contributing to public health, agriculture, and environment. It helps to identify emerging fungal allergens, track climate change impacts, and recognize conditions for disease outbreaks. The aim of this extramural and intramural study was to get knowledge about valuable information regarding identification, concentration and diversity of aeromycoflora in both indoor and outdoor environment to understand the cumulative aeromycoflora composition in different locations of Gorakhpur city.

## 2. Materials and methods

### 2.1. Selection of study area

Gorakhpur is a district in Uttar Pradesh, India, located at approximately 26.76°N latitude and 83.37°E longitude near Indo-Nepal border. It lies in subtropical region of country, characterized by hot summers (25°C–40°C), heavy monsoon rains (July–September), and cool winters (5°C–20°C) with frequent fog. The total eight study sites viz. Central library (CL), Deen Dayal Upadhyaya Gorakhpur University (DDUGU); Baba Raghav Das (BRD), medical college; All India Institute of Medical Sciences (AIIMS); Gorakhnath temple (GT); Botanical Garden (BG), DDUGU; Fertilizer factory (FF); Kushmi forest (KF) and Ramgarh tal/Nauka Vihar (RT/NV) in Gorakhpur district have been selected based on crowd mobility throughout day. CL, BRD, AIIMS and GT were selected to study indoor mycoflora from their central hall, outpatient department (OPD), OPD and outer hall respectively. The rest four sites i.e. BG, FF, KF and RT/NV were chosen to study outdoor mycoflora from their medicinal garden, urea loading area, forest and bank of RT/NV respectively.



**Fig. 1** Study sites in Gorakhpur district for aeromycobiota analysis

## 2.2. Determination of temperature and relative humidity of air

The temperature and relative humidity (RH) of study sites have been determined at the time of sampling by digital temperature and humidity meter (HTC HD-303) in morning (10-11 am) and late afternoon (4-5 pm).

## 2.3. Collection of air samples

Air samples were collected in the first week of October, 2023 on five different dates i.e. 02<sup>nd</sup>, 03<sup>rd</sup>, 05<sup>th</sup>, 06<sup>th</sup> and 7<sup>th</sup> October. During sampling, pre-poured Petri dishes with sterilized potato dextrose agar (PDA) medium prepared in 0.5% saline solution (to suppress Mucorales members) were exposed in air for 15 min. at study sites to trap the aeromycobiota. The samples were collected both in morning (10-11 am) and late afternoon (4-5 pm) by open plate technique (Menghare & Kayarkar, 2023). Ten poured Petri dishes were used to collect air samples in morning and late afternoon for each study sites separately. The inoculated Petri dishes with air samples were incubated ( $27 \pm 2^\circ\text{C}$ ) for seven days (Ghosh et al., 2011).

## 2.4. Identification of fungal species

The cultural and morphological characteristics of recovered fungal colonies were determined by their macroscopic and microscopic observations. The micro and macro morphology, as well as the surface and reverse colony colors on PDA media, were used to identify the fungal species (Ghosh et al., 2011). For microscopic study, fungi were stained and mounted in lactophenol cotton blue mixture. The fungi were identified up to genus and species level using manuals of fungi (Moubasher, 1993; Gilman, 2008; Mukerji & Manoharachary, 2010). The identified fungal colonies were purified and preserved on PDA slant at  $4 \pm 2^\circ\text{C}$ .

## 2.5. Statistical analysis

Temperature and relative humidity measurements were recorded five times, and the results are shown as mean  $\pm$  SE. The percent relative density and percent occurrence frequency was calculated as follows –

$$\text{Relative density (\%)} = \frac{\text{Number of isolates of a species}}{\text{Total no. of isolates of all species}} \times 100$$

$$\text{Occurrence frequency (\%)} = \frac{\text{Total number of isolates at one study site}}{\text{Total no. of isolates of all study sites}} \times 100$$

A one-way ANOVA was performed on the collected data of temperature and relative humidity. Tukey's multiple range tests were used to separate the means when the ANOVA was significant ( $p < 0.05$ ). Statistical software (SPSS 16.0; IBM, NY, USA) was used for data analysis.

### 3. Results

In the present study, average temperature and relative humidity of five different days i.e. 02<sup>nd</sup>, 03<sup>rd</sup>, 05<sup>th</sup>, 06<sup>th</sup> and 7<sup>th</sup> October, 2023 at eight different study sites (Fig. 1) were recorded both in morning (10-11 am) and late afternoon (4-5 pm). The lowest (21.68°C) and highest (24.90°C) average temperature in morning was recorded in KF and FF respectively. Whereas, in late afternoon, lowest (25.88°C) and highest (29.52°C) average temperature was found in KF and RT/NV respectively (Table 1). Similarly, lowest (54.76%) and highest (65.18%) mean relative humidity in morning was recorded in BRD and BG respectively. Whereas, in late afternoon, lowest (56.18%) and highest (70.08%) average relative humidity was found in BRD and KF respectively (Table 1).

**Table 1: Temperature and relative humidity of selected study sites**

S.No.	Study Sites		Coordinates	Temperature (°C)		RH (%)	
				Morning (10-11 am)	Late afternoon (4-5 pm)	Morning (10-11 am)	Late afternoon (4-5 pm)
1.	Indoor	Central library, DDUGU	26°44'54.7"N 83°22'52.8"E	24.04±0.09 <sup>bc</sup>	27.72±0.12 <sup>c</sup>	64.00±0.12 <sup>c</sup>	66.14±0.12 <sup>d</sup>
2.		BRD Medical College	26°48'44.3"N 83°23'59.2"E	24.58±0.16 <sup>cd</sup>	28.98±0.06 <sup>e</sup>	54.76±0.17 <sup>a</sup>	56.18±0.09 <sup>a</sup>
3.		AIIMS, Gorakhpur	26°44'49.9"N 83°25'07.3"E	23.44±0.14 <sup>b</sup>	27.06±0.07 <sup>b</sup>	60.28±0.20 <sup>c</sup>	62.02±0.13 <sup>b</sup>
4.		Gorakhnath Temple	26°46'22.7"N 83°21'30.4"E	24.40±0.12 <sup>cd</sup>	28.88±0.09 <sup>e</sup>	62.62±0.12 <sup>d</sup>	64.48±0.13 <sup>c</sup>
5.	Outdoor	Botanical Garden, DDUGU	26°44'45.3"N 83°22'55.0"E	23.60±0.18 <sup>b</sup>	28.26±0.12 <sup>d</sup>	65.18±0.14 <sup>f</sup>	68.54±0.18 <sup>e</sup>
6.		Fertilizer Factory	26°49'04.8"N 83°21'58.3"E	24.90±0.16 <sup>d</sup>	28.32±0.11 <sup>d</sup>	58.82±0.09 <sup>b</sup>	62.14±0.14 <sup>b</sup>
7.		Kushmi Forest	26°45'15.2"N 83°28'48.8"E	21.68±0.12 <sup>a</sup>	25.88±0.09 <sup>a</sup>	64.96±0.19 <sup>f</sup>	70.08±0.27 <sup>f</sup>
8.		Ramgarh Tal	26°43'23.9"N 83°24'06.3"E	23.98±0.11 <sup>bc</sup>	29.52±0.04 <sup>f</sup>	62.90±0.15 <sup>d</sup>	64.40±0.07 <sup>c</sup>

Values are mean ( $n = 5$ ) ± SE;  $P < 0.05$ . The means followed by same letter in the same column are not significantly different according to One-Way ANOVA and Tukey's multiple comparison tests

A total of 21 identified species belonging to 13 fungal genera were isolated excluding Mucorales, mycelia sterilia and unidentified fungi. The findings reveals that a total of 1506 and 2287 fungal colonies were recovered from air sampling of study sites during morning and late afternoon respectively (Table 2 & 3). The highest mycoflora occurrence frequency (%) was found in KF both in morning (19.46%) and late afternoon (22.60%) while, lowest in morning (6.30%) and late afternoon (7.08%) was determined in BRD and AIIMS (Table 2 & 3, Fig. 2). Collectively, during morning sampling, *Cladosporium* sp. exhibited highest relative density (19.85%) followed by *Aspergillus flavus* (17.13%), *Aspergillus niger* (16.73%) and *Penicillium* sp. (10.89%), whereas, both *Acremonium* sp. and *Colletotrichum* sp. (0.13%) having lowest relative density followed by *Alternaria atra* (0.20%), and *Alternaria longipes* (0.33%) (Table 2, Fig. 3). The late afternoon sampling reveals, *Cladosporium* sp. had highest relative density (22.39%)



followed by *Aspergillus. niger* (21.47%), *Aspergillus. flavus* (15.26%), *Aspergillus. fumigates* (9.49%) and *Penicillium* sp. (8.09%), whereas, *Colletotrichum* sp. (0.35%) having lowest relative density followed by *Alternaria longipes* (0.39%), *Acremonium* sp. (0.44%), *Rhizoctonia* sp. (0.48%) and *Alternaria atra* (0.52%) (Table 3, Fig. 3).

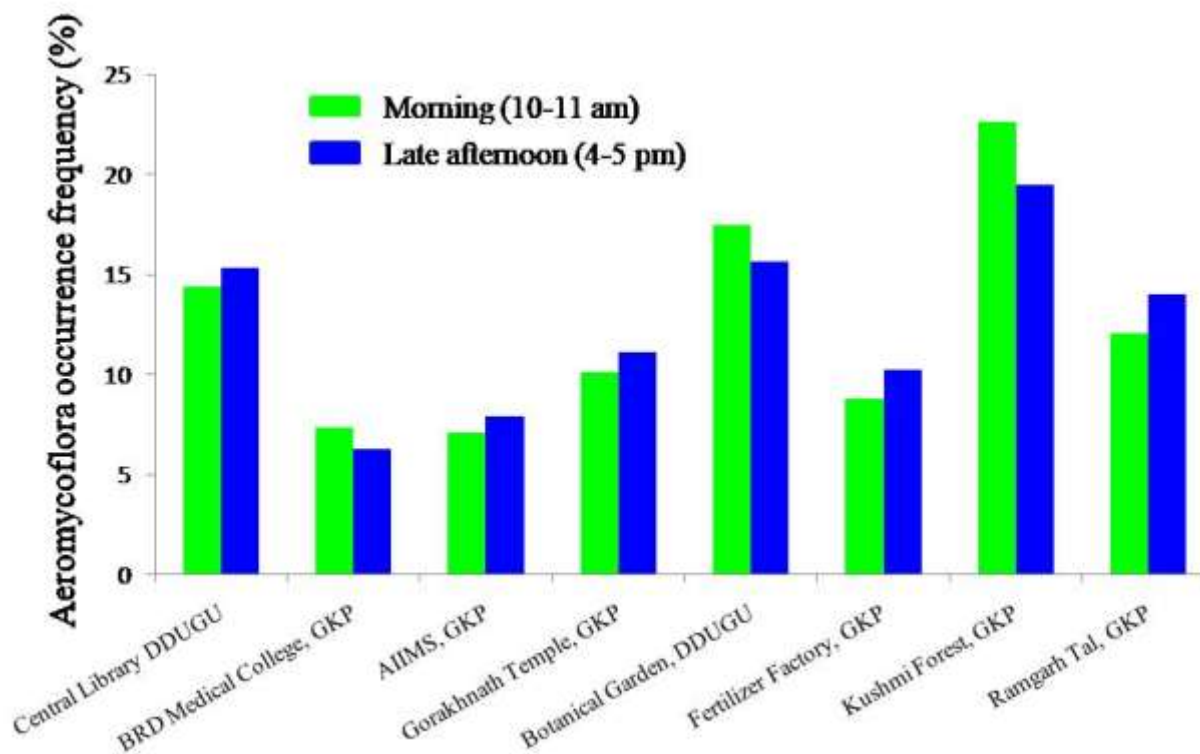


Fig. 2: Occurrence frequency (%) of aeromycobiota at different study sites

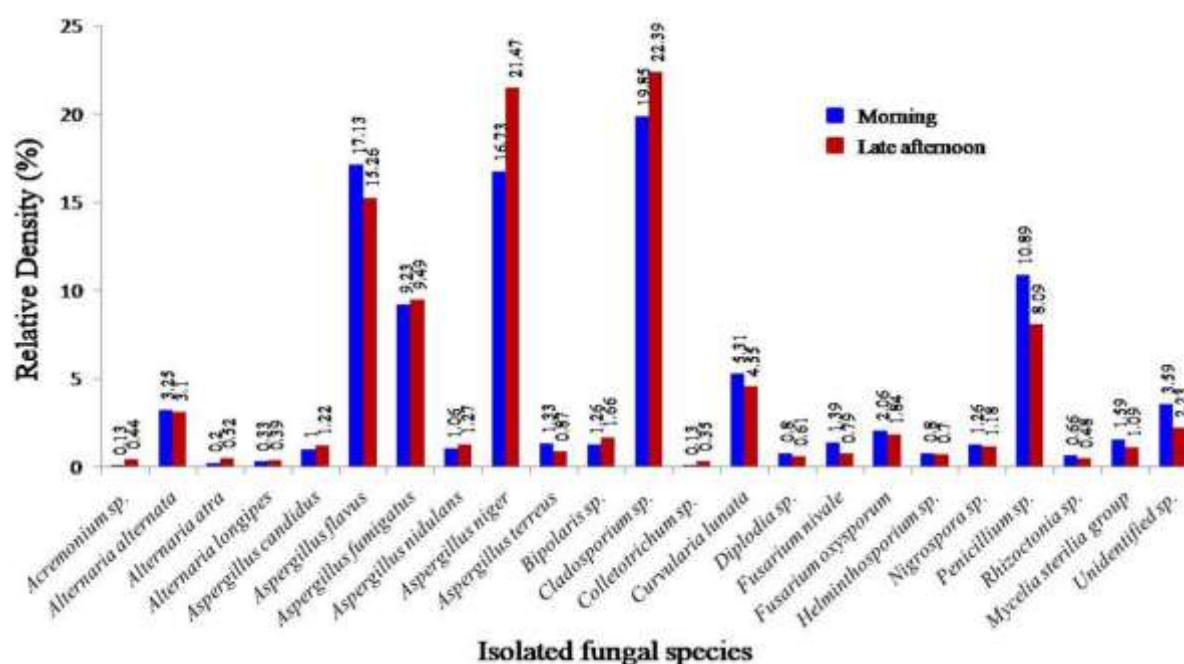


Fig. 3: Cumulative relative densities (%) of isolated aeromycobiota from different study sites

**Table 2: Aeromycobiota diversity in morning (10-11 am) at different experimental sites in Gorakhpur (GKP)**

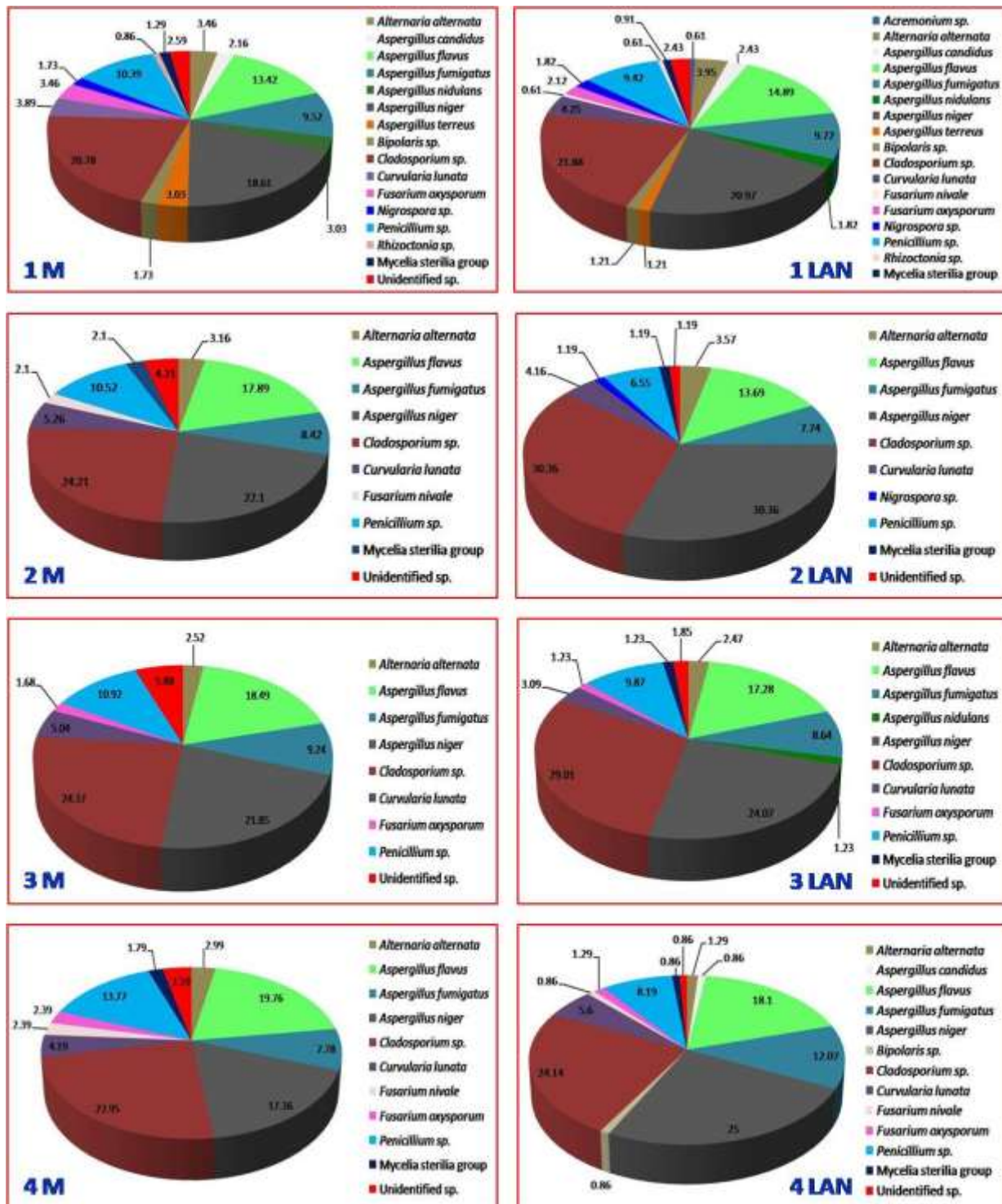
Fungal Species	Total number of fungal isolates								Total Isolates	Relative Density (%)
	Indoor Sites				Outdoor Sites					
	Centr al Librar y DDU GU	BRD Medical College, GKP	AIIMS, GKP	Gorakhnath Temple, GKP	Botanic al Garden , DDUG U	Fertili zer Facto ry, GKP	Kushmi Forest, GKP	Ra mgar h Tal, GK P		
<i>Acremonium</i> sp.	-	-	-	-	-	-	02	-	02	0.13
<i>Alternaria alternata</i>	08	03	03	05	07	05	14	04	49	3.25
<i>Alternaria atra</i>	-	-	-	-	-	-	03	-	03	0.20
<i>Alternaria longipes</i>	-	-	-	-	-	-	05	-	05	0.33
<i>Aspergillus candidus</i>	05	-	-	-	03	-	03	04	15	1.00
<i>Aspergillus flavus</i>	31	17	22	33	37	41	43	34	258	17.13
<i>Aspergillus fumigatus</i>	22	08	11	13	29	12	19	25	139	9.23
<i>Aspergillus nidulans</i>	07	-	-	-	05	-	-	04	16	1.06
<i>Aspergillus niger</i>	43	21	26	29	32	31	37	33	252	16.73
<i>Aspergillus terreus</i>	07	-	-	-	05	-	06	02	20	1.33
<i>Bipolaris</i> sp.	04	-	-	-	03	-	12	-	19	1.26
<i>Cladosporium</i> sp.	48	23	29	40	39	32	51	37	299	19.85
<i>Colletotrichum</i> sp.	-	-	-	-	-	-	02	-	02	0.13
<i>Curvularia lunata</i>	09	05	06	07	14	12	14	13	80	5.31
<i>Diplodia</i> sp.	-	-	-	-	02	-	08	02	12	0.80
<i>Fusarium nivale</i>	-	02	-	04	04	-	06	05	21	1.39
<i>Fusarium oxysporum</i>	08	-	02	04	06	02	05	04	31	2.06
<i>Helminthosporium</i> sp.	-	-	-	-	02	-	07	03	12	0.80
<i>Nigrospora</i> sp.	04	-	-	-	05	-	07	03	19	1.26
<i>Penicillium</i> sp.	24	10	13	23	31	11	28	24	164	10.89
<i>Rhizoctonia</i> sp.	02	-	-	-	02	-	04	02	10	0.66
<i>Mycelia sterilia</i> group	03	02	-	03	03	03	06	04	24	1.59
Unidentified sp.	06	04	07	06	07	05	11	08	54	3.59
Mucorales* (No. of Genera)	04*	03*	03*	02*	04*	03*	04*	04*	-	-
Total Isolates	231	95	119	167	236	154	293	211	1506	
Frequency of Occurrence (%)	15.34	6.30	7.90	11.09	15.67	10.23	19.46	14.01		

Mucorales\* including *Rhizopus* sp., *Mucor* sp., *Mortierella* sp., *Circinella* sp. and *Absidia corymbifera* were recovered but not included among the total isolates.

**Table 3: Aeromycobiota diversity in late afternoon (4-5 pm) at different experimental sites in Gorakhpur (GKP)**

Fungal Species	Total number of fungal isolates								Total Isolates	Relative Density (%)
	Indoor Sites				Outdoor Sites					
	Central Library DDUGU	BRD Medical College, GKP	AIIMS, GKP	Gorakhnath Temple, GKP	Botanical Garden, DDUGU	Fertilizer Factory, GKP	Kushmi Forest, GKP	Ramgarh Tal, GKP		
<i>Acremonium</i> sp.	02	-	-	-	01	-	07	-	10	0.44
<i>Alternaria alternata</i>	13	06	04	03	09	07	21	08	71	3.10
<i>Alternaria atra</i>	-	-	-	-	-	-	12	-	12	0.52
<i>Alternaria longipes</i>	-	-	-	-	02	-	05	02	09	0.39
<i>Aspergillus candidus</i>	08	-	-	02	05	03	04	06	28	1.22
<i>Aspergillus flavus</i>	49	23	28	42	57	39	65	46	349	15.26
<i>Aspergillus fumigatus</i>	32	13	14	28	45	19	39	27	217	9.49
<i>Aspergillus nidulans</i>	06	-	02	-	09	-	07	05	29	1.27
<i>Aspergillus niger</i>	69	51	39	58	82	42	96	54	491	21.47
<i>Aspergillus terreus</i>	04	-	-	-	03	02	08	03	20	0.87
<i>Bipolaris</i> sp.	04	-	-	02	04	-	25	03	38	1.66
<i>Cladosporium</i> sp.	72	51	47	56	78	64	86	58	512	22.39
<i>Colletotrichum</i> sp.	-	-	-	-	-	-	08	-	08	0.35
<i>Curvularia lunata</i>	14	07	05	13	17	09	27	12	104	4.55
<i>Diplodia</i> sp.	-	-	-	-	03	-	08	03	14	0.61
<i>Fusarium nivale</i>	02	-	-	02	04	-	07	03	18	0.79
<i>Fusarium oxysporum</i>	07	-	02	03	09	03	11	07	42	1.84
<i>Helminthosporium</i> sp.	-	-	-	-	05	-	09	02	16	0.70
<i>Nigrospora</i> sp.	06	02	-	-	11	-	05	03	27	1.18
<i>Penicillium</i> sp.	31	11	16	19	38	08	41	21	185	8.09
<i>Rhizoctonia</i> sp.	02	-	-	-	02	-	05	02	11	0.48
Mycelia sterilia group	03	02	02	02	04	03	07	02	25	1.09
Unidentified sp.	08	02	03	02	11	02	14	09	51	2.23
Mucorales* (No. of Genera)	04*	03*	03*	02*	05*	03*	05*	04*	-	-
Total Isolates	332	168	162	232	399	201	517	276	2287	
Frequency of Occurrence (%)	14.52	7.35	7.08	10.14	17.45	8.79	22.60	12.07		

Mucorales\* including *Rhizopus* sp., *Mucor* sp., *Mortierella* sp., *Circinella* sp. and *Absidia corymbifera* were recovered but not included among the total isolates.

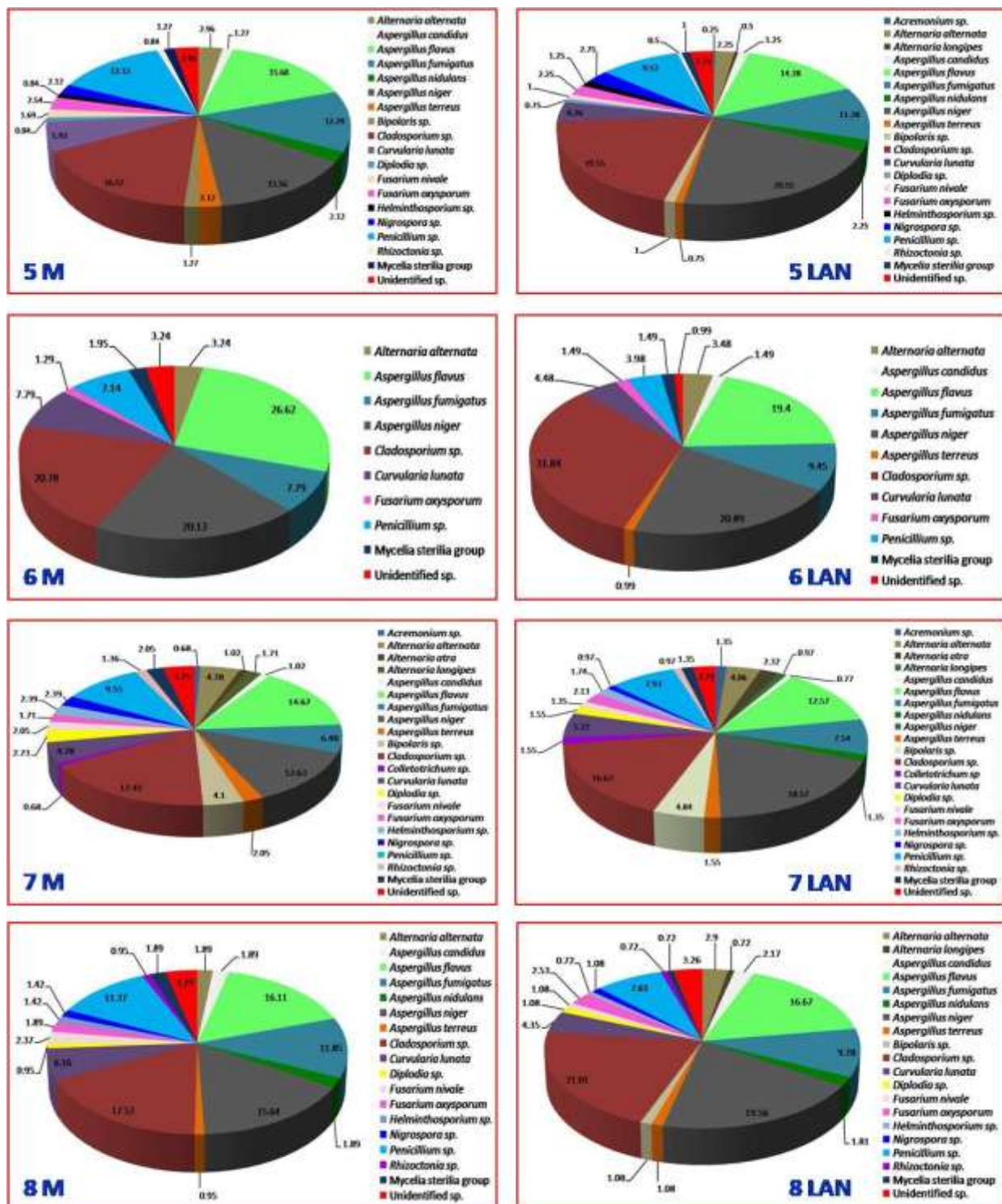


**M** – Morning (10-11 am); **LAN** – Late afternoon (4-5 pm)

**1**– Central Library, DDUGU; **2**– BRD Medical College; **3**– AIIMS, Gorakhpur; **4**– Gorakhnath Temple

**Fig. 4a: Relative densities (%) of isolated aeromycobiota from individual indoor study sites**





**M – Morning (10-11 am); LAN – Late afternoon (4-5 pm)**

**5- Botanical Garden, DDUGU; 6- Fertilizer Factory, GKP; 7- Kushmi Forest, GKP; 8- Ramgarh Tal, GKP**

**Fig. 4b: Relative densities (%) of isolated aeromycobiota from individual outdoor study sites**

Individually, in morning least aeromycological diversity (08 identified species) was found in both BRD and AIIMS, whereas, highest species diversity was recorded in KF (20 species) followed by BG (17 species). Similarly, in late afternoon, BRD exhibited least species diversity (08 species) followed by AIIMS (09 species) while, highest species diversity was again recorded in KF (21 species) followed by BG (19 species) (Table 1, 2; Fig. 4a, 4b). The distribution of aeromycoflora was found heterogeneous during study in CL, BG, KF and RT/NV. But other study sites were dominated by only few aeromycobiota. *A. niger*, *Cladosporium* sp. and

*A. flavus* occupied 74.41% of total aeromycoflora of BRD, similarly, 70.36 % aeromycoflora by *Cladosporium* sp., *A. niger* and *A. flavus* in AIIMS, 67.24 % by *A. niger*, *Cladosporium* sp. and *A. flavus* in GT, and 72.13 % by *Cladosporium* sp., *A. niger* and *A. flavus* in FF (Fig. 4a, 4b). The aeromycoflora at different study sites of Gorakhpur district was found dominated by *Aspergillus niger*, *Cladosporium* sp. and *Aspergillus flavus*.

#### 4. Discussion

Several literatures on aeromycoflora and their connection to associated environment have been published worldwide (Ianovici & Tudorica, 2009; Ababutain, 2013; Upadhyay et al., 2018; Karmakar et al., 2020). However, literature on aeromycoflora diversity in relation to their environment in Gorakhpur is still silent. The purpose of our study was to obtain the exact information about the fungal occurrences in air in relation to temperature and relative humidity fluctuation along with human disturbance in selected worthy sites.

In this investigation, total eight study sites were selected on the basis of their variable environment, location in Gorakhpur district and human disturbance. A modified gravitational plate method with movement at study sites was used to capture more aeromycoflora in lesser time. The overall lower number (1506) of fungal isolates in the morning in comparison to the late afternoon (2287) may be due to significant increase in temperature (Manikpuri et al., 2018), which possibly influence air velocity, bringing fungal spores along with it. Among study sites, the higher percent occurrence frequency in morning (19.46%) and late afternoon (22.60%) in KF supports the findings of earlier workers (Kunjam & Jadhav, 2020; Wagner et al., 2022; Ali et al., 2024), where forest soil, decomposing materials and phyllosphere fungi contributed in aeromycoflora diversity and lacking such conditions along with cleaning and sanitation processes, BRD and AIIMS exhibited least percent occurrence frequencies of aeromycoflora.

With significant raise in mean temperature and relative humidity from morning to late afternoon showed an increase in number of all fungal isolates from morning to late afternoon except *Colletotrichum* sp. and *Fusarium nivale*. The finding exhibited a direct correlation between increase in temperature and humidity with increase in number of fungal isolates (Crandall & Gilbert, 2017; Ajikah et al., 2023). Some aspergilli i.e. *Aspergillus niger*, *Aspergillus flavus*, *Aspergillus fumigates* along with *Cladosporium* sp. and *Penicillium* sp. were dominated in all samples in Gorakhpur due to their thermotolerance, rapid spore dispersal, and ability to thrive in diverse organic substrates. Their small lightweight conidia (spores), easily get dispersed by wind, ensuring their widespread presence in such climates. High humidity promotes their germination and growth on decaying plant material, soil, and indoor as well as outdoor surfaces, whereas warm environment enhance their sporulation and dissemination (Chiranjeevi et al., 2012; Crandall & Gilbert, 2017; Ajikah et al., 2023).

The total number of aeromycoflora isolates in indoor sites in morning (612) and late afternoon (794) was lesser than outdoor sites in morning (894) and late afternoon (1393). Such significant variation in indoor and outdoor aeromycoflora may occur due to differences in environmental conditions, sources of fungal spores, and human influence. Indoor aeromycoflora is primarily influenced by factors such as humidity, ventilation, building materials, and human activity. Whereas, outdoor aeromycoflora are influenced by temperature, humidity, soil disturbance, wind velocity and several anthropogenic factors (Ajikah et al., 2023). Seasonal fluctuations significantly affect outdoor aeromycoflora diversity, with higher concentrations in warm and humid conditions (Kakde et al., 2001).

Except KF and BG human hustle-bustle increased with passing time on rest of the other study sites. Along with other factors, human hustle-bustle also influence aeromycoflora population especially in indoor places i.e. CL, BRD, AIIMS and GT, outdoor mycoflora of settled dust may brought inside (Hicks et al., 2005). Most of the materials in the old libraries i.e. several literatures including books, journals, magazines, newspapers, wooden racks etc. are cellulosic and favors the growth of *Aspergillus* and other cellulolytic fungi (Díaz et al., 2020; Camargo Caicedo et al., 2023; Kujović et al., 2024), which could adversely affect student's health throughout the world (Valeriani et al., 2017). Aeromycoflora recovered from GT, FF and RT/NV was influenced by meteorological factors especially temperature, relative humidity and wind speed along with anthropogenic factors which influence aeromycoflora

diversity by affecting fungal growth, sporulation, and airborne spore dispersal. High humidity and warm temperatures promote fungal proliferation, while wind aids in the transportation of spores across regions. Pollution, agricultural activities, and construction activities also influence aeromycoflora diversity by modifying habitat conditions (Grinn-Gofroń et al., 2015; van Rhijn et al., 2021).

Several aerobiological evaluations have identified the presence of airborne fungi in indoor environments as one of the major health challenges experiencing India and other countries (Khan & Karuppayil, 2012; Singh & Mathur, 2012). However, earlier studies showed that fungus concentrations and the severity of the sickness in respiratory diseases, allergies, and infections especially in immunosensitive individuals were positively correlated (Rick et al., 2020). Through their interactions with internal human organs like as the lung, they raise the risk of respiratory disorders (Rick et al., 2020; Tiew et al., 2021; Palmieri et al., 2022). Additionally, airborne fungi also contribute to biodeterioration of various surfaces, artworks, and food products, leading to structural damage, along with allergies and mycotoxicosis in humans. Biomonitoring of airborne microfungi spores is a key to unlocking the information about sensitivity to bioaerosol in the environment of different locations, and our findings may be helpful in diagnosis and preventive measures of allergic illnesses caused by aeromycoflora.

## 5. Conclusion

The present study on the fluctuation of aeromycological diversity across different sites in Gorakhpur district, Uttar Pradesh, highlights the dynamic nature of airborne fungal populations influenced by temperature, humidity and human disturbance. Our analysis revealed a remarkable occurrence of several aeromycoflora with certain genera exhibiting higher prevalence in all the study sites including indoor and outdoor spaces. Some of them are well known as pathogens of plants and allergens to human, emphasizes the potential health risks associated with airborne fungal allergens and opportunistic pathogens. Many isolated fungal species are harmful and could cause respiratory disorders like asthma in visitors to libraries, temples, hospitals and other places. Therefore, suitable safeguards by identification and monitoring of aeromycoflora diversity is essential for assessing air quality, mitigating health hazards, and developing effective fungal control strategies.

## Acknowledgement

Authors are grateful to Head, Department of Botany, Deen Dayal Upadhyaya Gorakhpur University, Gorakhpur for providing laboratory facilities.

## Conflict of interest

The authors declare that they do not have any conflict of interest in this study.

## References

- Ababutain, I. M. (2013). Aeromycoflora of some eastern provinces of Saudi Arabia. *Indoor and Built Environment*, 22(2), 388-394. <https://doi.org/10.1177/1420326X114268>
- Adams, R. I., Miletto, M., Taylor, J. W., & Bruns, T. D. (2013). Dispersal in microbes: fungi in indoor air are dominated by outdoor air and show dispersal limitation at short distances. *The ISME Journal*, 7(7), 1262-1273.
- Ajikhah, L. B., Roffe, S. J., Neumann, F. H., Bamford, M. K., Esterhuizen, N., Berman, D., & Peter, J. (2023). Meteorological influences on airborne pollen and spores in Johannesburg (Gauteng), South Africa. *Aerobiologia*, 39(3), 363-388. <https://doi.org/10.1007/s10453-023-09799-2>
- Ali, E., Parbin, S., & Baruah, C. (2024). Analysis of indoor and outdoor aeromycoflora in two broiler farms of Barpeta, Assam. *International Journal of Biosciences*, 24(1), 28-38. <http://dx.doi.org/10.12692/ijb/24.1.28-38>



- Baxi, S. N., Portnoy, J. M., Larenas-Linnemann, D., Phipatanakul, W., Barnes, C., Baxi, S., & Williams, P. B. (2016). Exposure and health effects of fungi on humans. *The Journal of Allergy and Clinical Immunology: In Practice*, 4(3), 396-404. <https://doi.org/10.1016/j.jaip.2016.01.008>
- Camargo Caicedo, Y., Borja Pérez, H., Muñoz Fuentes, M., Vergara-Vásquez, E., & Vélez-Pereira, A. M. (2023). Assessment of fungal aerosols in a public library with natural ventilation. *Aerobiologia*, 39(1), 37-50. <https://doi.org/10.1007/s10453-022-09772-5>
- Castro e Silva, D. M., Marcusso, R. M. N., Barbosa, C. G. G., Gonçalves, F. L. T., & Cardoso, M. R. A. (2020). Air pollution and its impact on the concentration of airborne fungi in the megacity of São Paulo, Brazil. *Heliyon*, 6(10), e05065. <https://doi.org/10.1016/j.heliyon.2020.e05065>
- Chiranjeevi, T., Rani, G., Chandel, A. K., Sekhar, P. V., Prakasham, R. S., & Addepally, U. (2012). Optimization of holocellulolytic enzymes production by *Cladosporium cladosporioides* using taguchi-L'16 orthogonal array. *Journal of Biobased Materials and Bioenergy*, 6(2), 148-157.
- Crandall, S. G., & Gilbert, G. S. (2017). Meteorological factors associated with abundance of airborne fungal spores over natural vegetation. *Atmospheric Environment*, 162(8), 87-99. <https://doi.org/10.1016/j.atmosenv.2017.05.018>
- Díaz, G. V., Coniglio, R. O., Chungara, C. I., Zapata, P. D., Villalba, L. L., & Fonseca, M. I. (2020). *Aspergillus niger* LBM 134 isolated from rotten wood and its potential cellulolytic ability. *Mycology*, 12(3), 160-173. <https://doi.org/10.1080/21501203.2020.1823509>
- Durugbo, U. E., Adesanya, O. O., Adeleke, O., Adetutu, S., Tioluwani, A., Ayobami, O., & Adedotun, A. O. (2023). Distribution patterns of airborne bacteria and fungi in a teaching, public and private hospital in southwestern Nigeria. *Aerobiologia*, 39(4), 393-414. <https://doi.org/10.1007/s10453-023-09795-6>
- Estillore, A. D., Trueblood, J. V., & Grassian, V. H. (2016). Atmospheric chemistry of bioaerosols: heterogeneous and multiphase reactions with atmospheric oxidants and other trace gases. *Chemical Science*, 7(11), 6604-6616. <https://doi.org/10.1039/C6SC02353C>
- Ghosh, D., Dhar, P., Das, A. K., & Uddin, N. (2011). Identification and distribution of aeromycoflora in the indoor environment of Shyambazar Metro-Railway Station, Kolkata, India. *African Journal of Microbiology Research*, 5(31), 5569-5574.
- Gilman, J. C. (1998). *A manual of soil fungi*. Daya Books, New Delhi.
- Grinn-Gofroń, A., & Bosiacka, B. (2015). Effects of meteorological factors on the composition of selected fungal spores in the air. *Aerobiologia*, 31(1), 63-72. <https://doi.org/10.1007/s10453-014-9347-1>
- Grinn-Gofroń, A., Bosiacka, B., Bednarz, A., & Wolski, T. (2018). A comparative study of hourly and daily relationships between selected meteorological parameters and airborne fungal spore composition. *Aerobiologia*, 34(1), 45-54. <https://doi.org/10.1007/s10453-017-9493-3>
- Hicks, J. B., Lu, E. T., De Guzman, R., & Weingart, M. (2005). Fungal types and concentrations from settled dust in normal residences. *Journal of Occupational and Environmental Hygiene*, 2(10), 481-492.
- Ianovici, N., & Tudorica, D. (2009). Aeromycoflora in outdoor environment of Timisoara City (Romania). *Notulae Scientia Biologicae*, 1(1), 21-28. <https://doi.org/10.15835/nsb113446>
- Kakde, U., B., Kakde, H. U., & Saoji, A. (2001). Seasonal variation of fungal propagules in a fruit market environment, Nagpur (India). *Aerobiologia*, 17(2), 177-182. <https://doi.org/10.1023/A:1010849522964>
- Karmakar, B., SenGupta, K., Kaur, A., Roy, A., & Bhattacharya, S. G. (2020). Fungal bio-aerosol in multiple micro-environments from eastern India: source, distribution, and health hazards. *SN Applied Sciences*, 2(4), 1-14. <https://doi.org/10.1007/s42452-020-2323-1>
- Khan, A. H., & Karuppaiyil, S. M. (2012). Fungal pollution of indoor environments and its management. *Saudi Journal of Biological Sciences*, 19(4), 405-426.



- Kim, K. H., Kabir, E., & Jahan, S. A. (2018). Airborne bioaerosols and their impact on human health. *Journal of Environmental Sciences*, 67(5), 23-35. <https://doi.org/10.1016/j.jes.2017.08.027>
- Kochar, S., Ahlawat, M., Dahiya, P., & Chaudhary, D. (2014). Assessment of allergenicity to fungal allergens of Rohtak city, Haryana, India. *Allergy & Rhinology*, 5(2), e56-e65. <https://doi.org/10.2500/ar.2014.5.0088>
- Kujović, A., Gostinčar, C., Kavkler, K., Govedić, N., Gunde-Cimerman, N., & Zalar, P. (2024). Degradation potential of xerophilic and xerotolerant fungi contaminating historic canvas paintings. *Journal of Fungi*, 10(1), 76. <https://doi.org/10.3390/jof10010076>
- Kunjam, S., Jadhav, S.K. (2020). Diversity of soil and leaf surface mycoflora: A source of aeromycoflora. *Indian Journal of Aerobiology*. 3(1), 9–16.
- Letovsky, S., Robinson, M., Kwong, K., Liu, A. H., Sullivan, A., & Valcour, A. (2024). Assessing the contributions of phylogenetic and environmental determinants of allergic cosensitization to fungi in humans. *Annals of Allergy, Asthma & Immunology*, 132(2), 208-215. <https://doi.org/10.1016/j.anai.2023.10.016>
- Madsen, A. M., Hansen, V. M., Meyling, N. V., & Eilenberg, J. (2007). Human exposure to airborne fungi from genera used as biocontrol agents in plant production. *Annals of Agricultural and Environmental Medicine*, 14, 5-24.
- Manikpuri, M., Tiwari, K. L., & Tiwary, B. N. (2018). Effect of seasonal variation on aeromycoflora of Bilaspur, Chhattisgarh, involved in allergic reactions. *Aerobiologia*, 34, 119-126. <https://doi.org/10.1007/s10453-017-9500-8>
- Menghare V., & Kayarkar A. (2023). Seasonal Variation in Aeromycoflora of Satpuda Botanical Garden, Nagpur (M. S.), India. *International Journal for Multidisciplinary Research*. 5(6), 01-08. <https://doi.org/10.36948/ijfmr.2023.v05i06.10060>
- Moubasher, A. H. (1993). *Soil Fungi in Qatar and Other Arab Countries*. The Centre of Scientific and Applied Research, University of Qatar.
- Mukerji, K. G., & Manoharachary, C. (2010). *Taxonomy and ecology of Indian fungi*. IK International Pvt Ltd.
- Niu, M., Hu, W., Cheng, B., Wu, L., Ren, L., Deng, J., & Fu, P. (2021). Influence of rainfall on fungal aerobiota in the urban atmosphere over Tianjin, China: A case study. *Atmospheric Environment: X*, 12, 100137. <https://doi.org/10.1016/j.aeaoa.2021.100137>
- Oliveira, M., Oliveira, D., Lisboa, C., Boechat, J. L., & Delgado, L. (2023). Clinical manifestations of human exposure to fungi. *Journal of Fungi*, 9(3), 381. <https://doi.org/10.3390/jof9030381>
- Palmieri, F., Koutsokera, A., Bernasconi, E., Junier, P., von Garnier, C., & Ubags, N. (2022). Recent advances in fungal infections: from lung ecology to therapeutic strategies with a focus on *Aspergillus* spp. *Frontiers in Medicine*, 9, 832510. <https://doi.org/10.3389/fmed.2022.832510>
- Park, S. H., Kang, B. R., Kim, J., Lee, Y., Nam, H. S., & Lee, T. K. (2024). Enhanced soil fertility and carbon dynamics in organic farming systems: The role of arbuscular mycorrhizal fungal abundance. *Journal of Fungi*, 10(9), 598. <https://doi.org/10.3390/jof10090598>
- Pavan, R., & Manjunath, K. (2014). Qualitative analysis of indoor and outdoor airborne fungi in cowshed. *Journal of Mycology*, 1-8. <https://doi.org/10.1155/2014/985921>
- Prussin, A. J., & Marr, L. C. (2015). Sources of airborne microorganisms in the built environment. *Microbiome*, 3, 1-10. <https://doi.org/10.1186/s40168-015-0144-z>
- Pusz, W., Król, M., & Zwijacz-Kozica, T. (2018). Airborne fungi as indicators of ecosystem disturbance: an example from selected Tatra Mountains caves (Poland). *Aerobiologia*, 34(1), 111-118. <https://doi.org/10.1007/s10453-017-9498-y>
- Rick, E. M., Woolnough, K., Pashley, C. H., & Wardlaw, A. J. (2016). Allergic fungal airway disease. *Journal of Investigational Allergology & Clinical Immunology*, 26(6), 344-354. <https://doi.org/10.18176/jiaci.0122>
- Rick, E. M., Woolnough, K. F., Seear, P. J., Fairs, A., Satchwell, J., Richardson, M., & Pashley, C. H. (2020). The airway fungal microbiome in asthma. *Clinical & Experimental Allergy*, 50(12), 1325-1341. <https://doi.org/10.1111/cea.13722>
- Singh, A. B., & Mathur, C. (2012). An aerobiological perspective in allergy and asthma. *Asia Pacific Allergy*, 2(3), 210-222.

- Srivastava, N., Elgorban, A. M., Mishra, P. K., Marraiki, N., Alharbi, A. M., Ahmad, I., & Gupta, V. K. (2020). Enhance production of fungal cellulase cocktail using cellulosic waste. *Environmental Technology & Innovation*, 19, 100949. <https://doi.org/10.1016/j.eti.2020.100949>
- Tastassa, A. C., Sharaby, Y., & Lang-Yona, N. (2024). Aeromicrobiology: a global review of the cycling and relationships of bioaerosols with the atmosphere. *Science of the Total Environment*, 912(1), 168478. <https://doi.org/10.1016/j.scitotenv.2023.168478>
- Tiew, P. Y., Mac Aogáin, M., Ter, S. K., Aliberti, S., Chalmers, J. D., & Chotirmall, S. H. (2021). Respiratory mycoses in COPD and bronchiectasis. *Mycopathologia*, 186, 623-638. <https://doi.org/10.1007/s11046-021-00539-z>
- Upadhyay, H., Banik, D., Siddique, A., & Kumar, A. (2018). Aeromycoflora of fruit and vegetables market environment and their proper management towards a sustainable environment. *Plant Archives*, 18(2), 1851-1854.
- Vaksmaa, A., Guerrero-Cruz, S., Ghosh, P., Zeghal, E., Hernando-Morales, V., & Niemann, H. (2023). Role of fungi in bioremediation of emerging pollutants. *Frontiers in Marine Science*, 10, 1070905. <https://doi.org/10.3389/fmars.2023.1070905>
- Valeriani, F., Cianfanelli, C., Gianfranceschi, G., Santucci, S., Spica, V. R., & Mucci, N. (2017). Monitoring biodiversity in libraries: A pilot study and perspectives for indoor air quality. *Journal of Preventive Medicine and Hygiene*, 58(3), E238-E251.
- Van Rhijn, N., Coleman, J., Collier, L., Moore, C., Richardson, M. D., Bright-Thomas, R. J., & Jones, A. M. (2021). Meteorological factors influence the presence of fungi in the air; A 14-month surveillance study at an adult Cystic Fibrosis center. *Frontiers in Cellular and Infection Microbiology*, 11, 759944. <https://doi.org/10.3389/fcimb.2021.759944>
- Vijayalakshmi, P., Usharani, V., & Reddy, M. K. (2020). Studies on aeromycoflora in north coastal Andhra Pradesh, India. *Annals of Biology*, 36(3), 382-388.
- Wagner, R., Montoya, L., Gao, C., Head, J. R., Remais, J., & Taylor, J. W. (2022). The air mycobiome is decoupled from the soil mycobiome in the California San Joaquin Valley. *Molecular Ecology*, 31(19), 4962-4978. <https://doi.org/10.1111/mec.16640>