

RESEARCH ON TOPICAL GEL CONTAINING SOLID LIPID NANOPARTICLES LOADED WITH ACYCLOVIR

VIPUL SAINI^{*1}, RITU SAINI¹, JITENDRA KUMAR²

1. SMT. TARAWATI INSTITUTE OF BIOMEDICAL & ALLIED SCINCES, ROORKEE, UTTARAKHAND, INDIA 2. IBMERDOP MANGALAYATAN UNIVERSITY, ALIGARH, UTTAR PRADESH, INDIA

ABSTRACT

For extended therapeutic effect in the treatment of viral infections, new topical pharmacological alternatives are urgently required. In order to produce an efficient topical Acyclovir solid lipid nanoparticles (SLN) gel formulation with long-term therapeutic potential against tropical viral infections, bioavailability hurdles of Acyclovir were overcome. Acyclovir is a topical antiviral medication used to treat viral infections. Stearic acid and poloxamer 188 were used in the solvent diffusion approach to create the SLN for acyclovir. For the leading moiety's veracity, preformulation investigations were carried out. The generated SLN and gel formulations were then put through physicochemical testing, in-vitro drug release profiles, and kinetics studies. Following that, successful FTIR spectroscopy and scanning electron microscopy of the improved formulation were performed. The findings suggest that SLN F6 has a strong entrapment efficacy, with the greatest entrapment of 92.13%0.975. In terms of particle size, size distribution, and zeta potential, SLN show a mean particle diameter of about 344.3 nm, a unimodal size distribution, a polydispersity index of 0.168, an intercept value of 0.98 with 92% peak intensity, and a zeta potential of around 18.8 mV. Moreover, compared to other formulations, G3 gel exhibits a greater entrapment efficacy of 91.39%0.187. The G3 gel with 1.5% carbopol 934 w/v exhibits a sustained drug release profile with 79.57%0.213 of the drug release even after 24 hours. It is concluded that the Acyclovir loaded SLN based gel formulation containing carbopol 934 1.5% w/v is suitable for topical application and may show a much better result of anti-viral activity.

KEYWORDS: Solid lipid nanoparticles loaded Gel, Drug Content, pH of the Gel, In-vitrodrug release study

INTRODUCTION

Solid lipid nanoparticles (SLNs) are a cutting-edge, revolutionary drug delivery technology in the pharmaceutical industry today (NDDS). Typical colloidal carriers including polymeric and micro, liposome emulsions, and nanoparticles are represented by the SLN, which was first identified in 1991. The contemporary SLN technique is linked to improved drug penetration, a robust release profile, and targeted drug administration with great physical stability and low degradability, among other things. 58, 60 The benefits of nanoparticles with diameters ranging from 10 to 1000 nm on improving medication bioavailability seem encouraging. In the era of colloidal drug carrier systems, which produces an alternative particle in the field of NDDS, formulation hyphenated with SLN is a crucial consideration.

Solid Lipid Nanoparticles

Solid lipid nanoparticles (SLNs) range in size from 50 to 1000 nm and are colloidal drug carrier systems composed of solid lipid diffuse in aqueous surface active solution.

Due to qualities including good tolerance, physical stability, biodegradability, high bioavailability, protection of relevant pharmaceuticals from decay, ease of preparation, and low toxicity, ²¹ SLN offer a better alternative to polymeric systems, colloidal systems, and other NDDS. ^{38, 43, 67} Furthermore, the production process can be changed to release the required drug, prevent drug deterioration, and stay away from organic solvents. This broad adaptability can be crucial for the marketing of new items. A carrier method for enhanced drug delivery, SLN is an intriguing quality. ¹

Structure of solid lipid nanoparticles



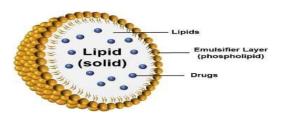


Figure 1: Structure of solid lipid nanoparticles. ³⁶

| Advantages of SLNs | |
|---|--------------|
| | Increase the |
| medications' stability. | |
| | Outstanding |
| biocompatibility. | - |
| 3. 5 | Steer clear |
| of organic solvents. ⁷⁴ | |
| 4. I | Long-term |
| stability, # ⁴ . | |
| 5. Increase trapped bioactive chemicals' bioavailability. | |
| 6. Sterilization and scaling up are very simple. | |
| 7. Manage and plan drug release. ²¹ | |
| 8. Minimal toxin. | |
| 9. Biocompatible and biodegradable | |
| Disadvantages of SLNs | |
| 1. Poor drug loading capacity. ⁶⁸ | |
| 2. Less compatibility for hydrophilic drugs | |
| 3. Unsure gelation tendency. | |
| Gels | |
| A "semisolid system where the liquid phase is blocked in a polymer matrix that causes a high degr | ree of |
| physical and chemical cross-linking" is what is meant by a gel. | |
| Properties of gels: | |
| 1 Gelling agents are employed in cosmetic and medicinal formulations. | |
| 2. It should be secure, inert, and free from interactions with other formulation elements | |
| 3. The addition of a gelling agent to the formulation will provide a logical solid-like co | • |
| during storage that can easily be broken down when shear forces are produced by squeezing the co | ontainer, |
| vortexing the bottle, or applying the formulation topically. | |
| 4. Using anti-microbials should make it possible to prevent microbial strike. | |

Using anti-microbials should ma
To avoid sloppiness.

It must to be sterile, much as ophthalmic gel

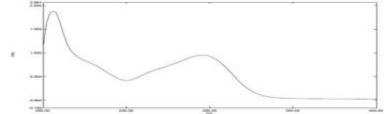
Preformulation study of drug

Melting Point: (*IP., 2007*) Melting point of Acyclovir was found to be $256.6 \pm 1.15^{\circ}$ C with decomposition.

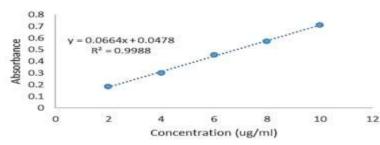
Determination of the absorption maximum of Acyclovir inethanol

The potential drug absorption was determined according to protocol using the highest amount of acyclovir that could be absorbed at 299 nm against a concentration of 2-10 g/ml. It was discovered that the regression

equation and coefficient were 0.0664x - 0.0478 and 0.998, respectively. Acyclovir absorption maximum determination and technique validation are related goals for qualitative and quantitative analysis.⁴⁸



Absorption maxima of Acyclovir in methenol



Regression coefficient of Acyclovir against concentration

Figure 3: Absorption maxima of Acyclovir and regression coefficient against the different concentration of Acyclovir (µg/ml)

Acyclovir's physicochemical studies were carried out to assess the drug's physicochemical characteristics. studies to assess the compatibility of acyclovir with hydrophilic and lipophilic compounds. The results demonstrate that Acyclovir has a low solubility potential in water, where it was found to be 0.00585 0.293 mg/ml, and in stearic acid, prectrol, and dynasan 114, where it was found to be 23.754 0.47, 18.314 0.85, and 22.875 0.32 mg/ml, respectively. Moreover, Acyclovir's non-aqueous solubility in n-octanol was 17.984 0.52 mg/ml. Acyclovir's log10P value in stearic acid, prectrol, dynasan 114, and n-octanol was simultaneously 3.98, 3.30, 3.87, and 3.65.

For a better compatibility examination of the leading moiety before and after formulation, acyclovir and stearic acid underwent FTIR analysis. Figure 4 and Table 1 show the FTIR spectra of acyclovir. Major IR absorption peaks of Acyclovir were observed at 2979.43 cm-1 (C-H stretch), 2198.82 cm-1 (C-N stretch), 1554.35 cm-1 (C-H aromatics stretch), 1471.35 cm-1 (C=C-C aromatic ring stretch), 820.41 cm-1 (para C-H distribution), and 759.46 cm-1 (C-Cl stretch). The main peaks that were discovered proved the Acyclovir's legitimacy and purity as being similar to the report that was used as a reference. ⁵²

| Table 1: FTIR interpretation of Acyclovir | | | | |
|---|------------------------------|---------------------------|--|--|
| S.No. | Wave number (cm ⁻ | Inference | | |
| 1 | 3441.02 | N-H stretching | | |
| 2 | 3308.74 | O-H stretching | | |
| 3 | 3099.14 | C-H stretching | | |
| 4 | 1717.21 | C=O stretching | | |
| 5 | 1541.50 | C=C and C=N stretching | | |
| 6 | 1308.75 | -CH2 Wagging and twisting | | |
| 7 | 1216.74 | Aryl alkyl ether | | |
| 8 | 1105.79 | C-O stretching | | |
| 9 | 1083.93 | C-O-C stretching | | |

| Similar to | the repe | i t tilut | was use | | |
|------------|----------|-----------|----------|--------|----------|
| Tabla 1 | . FTID | intonn | notation | n of l | avalarin |

nternational Journal of Scientific Research in Engineering and Management (IJSREM) Volume: 07 Issue: 05 | May - 2023

SJIF 2023: 8.176

ISSN: 2582-3930

Figure 4 FTIR spectrum of Acyclovir

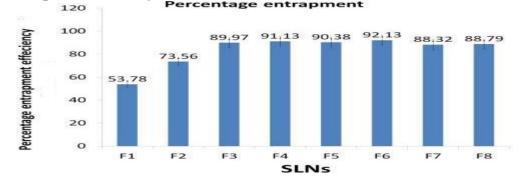
Development of the method for Acyclovir loaded solid lipidnanoparticles (SLN)

The technique involves a variety of modified nano-precipitation techniques, such as cooling sonication probe and nano-precipitation, for optimising SLN in relation to EE of acyclovir. 4°C and 25°C are the segmental temperature controls. A 4°C aqueous phase is instantly added to, and the resultant hyphenation with an organic phase causes a rapid precipitate. To achieve homogeneity, the temperature was adjusted throughout the early stages of nano-precipitation. By reducing bigger crystal size and aggregation of milled beads, highpressure homogenization helps to achieve uniform homogeneity.⁸⁷ Moreover, stearic acid and poloxomer 188 (w/v) concentrations were alternately changed from 0.5 to 2% while the technique was being optimised for SLN. Following the successful coding of all produced groups of SLN, the percentage of the active moiety that was captured was measured spectrophotometrically at 299 nm. Statistics were used to analyse the collected data. Acyclovir-high SLN are selected as the best SLN and are put through additional testing. Table 2:

| Different concentration of leading reagents for the formation of SLN | | | | |
|--|------------------|----------------------|----------------------|--|
| SLN code | Acyclovir %(w/v) | Stearic acid % (w/v) | Poloxomer 188 %(w/v) | |
| F1 | 1 | 0.5 | 1 | |
| F2 | 1 | 0.7 | 1 | |
| F3 | 1 | 1 | 1 | |
| F4 | 1 | 2 | 1 | |
| F5 | 1 | 1 | 0.5 | |
| F6 | 1 | 1 | 0.7 | |
| F7 | 1 | 1 | 1.5 | |
| F8 | 1 | 1 | 2 | |

| : Preparation of different Acyclovir solid lipid nanoparticle |
|---|
|---|

EVALUATION OF SLN Evaluation of entrapment efficiency of SLN







Acyclovir was initially evaluated spectroscopically and physicochemically in pre-formulation research. Percentage EE of Acyclovir was established following the successful production of various batches of nanoparticles. EE assessed spectrophotometrically at 299 nm as a percentage. The subsequent results show that SLN F6 and SLN F1 have the highest and lowest percentages of EE of Acyclovir loaded SLN, respectively, by 92.13%0.975 and 53.78%1.052 w/w. A study quoted by Ige et al. revealed the maximum% EE was between 90 and 95% w/w. 39 As a result, SLN F6 was chosen as an optimal SLN based on the percentage of drug entrapment, and evaluations of its physicochemical characteristics and gel formation followed. Graphical representations of the percentage of all SLN groups that were drug-entrapped.

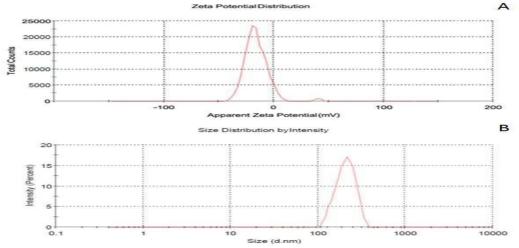
Physicochemical property

Based on their physicochemical properties, including colour, odour, pH stability, and water solubility, the SLN F6 were assessed. Physical and chemical analyses show that SLN is sufficiently more soluble in water than Acyclovir, having a white translucent colour, homogenous and uniform texture, fragrant flavour, and greater stability at pH.

Zeta potential and particle size and size distribution identification

Acyclovir SLN's zeta potential and particle size were successfully measured using the nano ZS90 zetasizer device. One of the key variables used to predict the physical stability of nanoparticles is zeta potential. A high zeta potential value is necessary for the stability of a nanoparticle system and indicates that a nanosystem will be more stable since it will be able to produce a dissuasive force between the nanoparticles. According to Fig. 6, SLN has a high zeta potential of 18.8 mV and indicates a highly stable nanosystem. SLN revealed a mean particle diameter of 344.3 nm, a unimodal size distribution, a polydispersity index (PDI) of 0.168, an intercept value of 0.98, and a peak intensity of 92%. When the PDI value is less than 0.5, it shows the dispersion factor with little nanoparticle agglomeration.⁸⁸

Figure 6 zeta potential, particle size and size distribution of Acyclovir SLN F6



Optical microscopy

Optical microscopy of the improved preparation, SLN F6, characterised with a digital light optical microscope at a 100x magnification, and observation reveal that Acyclovir SLN is successfully localised with homogeneous and uniform texture inside SLN dispersion. It claims that the only particles that could be seen clearly against the microscope resolution power were those whose mean diameter was larger than 2.5 m. Furthermore, no self-assembled structures have been seen in SLN preparation. When using optical microscopy, micellar structures were not seen. Figure 7 displays optical microscopy pictures of SLN F6 with Acyclovir put on it.

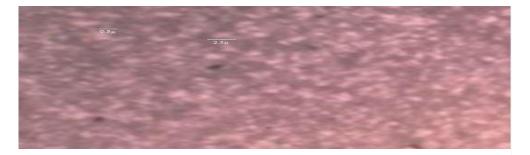


Figure 8 Drug-excipient comparability study by FTIR

To ascertain any potential interactions between the medicine and drug additives, FTIR analysis of SLN F6 was conducted. The main absorption peaks of acyclovir, according to spectral data, are at 2955.75 cm-1 for C-H stretching, 2523 and 2647 cm-1 for S-H stretching, 2201.52 cm-1 for CN stretching, 1556.90 cm-1 for C=N stretching, 1471.88 cm-1 for C=C aromatic ring stretching, and 720.33 and 1101.29 cm-1 for C-Cl stretching. Although the main stearic acid absorption peaks were discovered at 2914.97 cm-1 and 2848.05 cm-1 in the high-frequency range, attributed to the asymmetric and symmetric stretching vibrations of the -CH2- band, respectively, and at 1698.03 cm-1 for the -COOH stretching in the low-frequency zone. After the successful construction of the SLN, there have been no further alterations to Acyclovir, according to a spectral analysis of the optimised SLN. The reported referenced values are strongly supported by spectral data. ⁵²

| Characteristics Peaks | Reported (cm ⁻¹) | Observed(cm ⁻¹) |
|------------------------------------|------------------------------|-----------------------------|
| $-\mathbf{C} = \mathbf{O}$ stretch | 1650 - 1850 | 1717.21 |
| -COOH Stretch | 1600 - 1900 | 1771.95 |
| -NH amine stretch | 3150 - 3300 | 3441.02 |
| OH hydroxy stretch | 3150 - 3650 | 3308.74 |
| C-Cl stretch | 550 - 850 | 609.29 |

Table 2: FTIR interpretation of SLN F6

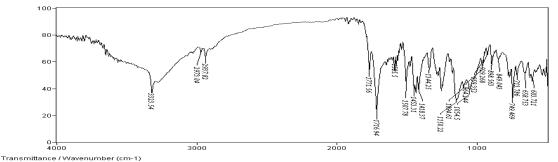


Figure 9: FTIR spectra of SLN F6

Optimization and evaluation of SLN gel

The preparation of the topical gel containing SLN loaded with Acyclovir was accomplished utilising the stirring method and carbopol 934 as the gelling agent. Unpretentious and reliable preparation methods for several SLN gels were discovered. First, the effectiveness of the four different SLN gel preparations, designated as G1, G2, G3, and G4, was assessed in order to measure the percentage of Acyclovir that was entrapped using spectrophotometry at 299 nm. The data obtained reveals that SLN G3 with 1.5% carbopol w/w has the highest drug entrapment percentage (91.39% 0.187). After that, the improved formulation is further assessed for physiochemical characteristics like spreadability, viscosity, pH, and appearance. Results show that the G3 gel's viscosity was determined to be 369cP, which is comparable to the gel viscosity reported by Jana et al. Additionally, the pH of the gel was determined to be 6.120.255.41 In the spreadability

evaluation, the spreadability factor of the prepared SLN gel was determined to be 4.5, and it is stated that the prepared gel produces excellent spreadability as an ideal topical formulation. From the perspective of patient compliance, spreadability is one of any topical formulation's crucial physical qualities. ¹⁸



Figure 10: visual appearance of SLN G3 gel

In vitro drug release and kinetics study

It is routine practise to compare release profiles and predict release mechanisms using statistical models. Drug in-vitro release profile was carried out in a buffer system utilising the dialysis bag method for 24 hours. As shown in Figure 15 and Table 10, the percentages of Acyclovir that desolvate from SLN rise proportionately with time. A developed SLN is capable of releasing drugs in a controlled manner, according to evidence from release profiles. Most SLN forms use homogenous drug trapping throughout systems to explain the leading moiety's slow release.⁶² According to Ekambaram et al., a regulated drug desolvation profile can be achieved when the drug is evenly distributed throughout the lipid matrix. Due to its larger HLB value than Cremophor RH 40, Poloxamer 407 has more efficacy against the rate of drug release from SLN.20 Furthermore, Poloxamer 407 has a high external spreadability, which lessens the impacts of the interfacial tension between SLN and the dissolution medium. Moreover, it speeds up drug disintegration and decreases the buildup of drug particles. Moreover, the lipid mass in SLN can improve the potency of the drug's desolvation and govern the size of nanoparticles. Drug release is delayed due to a lengthened effect due to the thickness of the lipid around the nanoparticle.¹⁷

| Sr. no. | Time i hours | in Percentage release of G3 | drug Percentage drug releaseof control gel |
|--------------|-----------------|--------------------------------|---|
| 1 | 0 | 0 | 0 |
| 2 | 0.25 | 7.375±0.153 | 1.923±0.011 |
| 3 | 0.5 | 14.002±0.185 | 2.052 ± 0.155 |
| 4 u | 1 | 22.064±0.102 | 3.042 ± 0.158 |
| 5 ea | 2 | 32.289±0.173 | 3.182 ± 0.162 |
| 8 9 | | 40.622±0.165 | 5.094 ± 0.122 |
| 7 a n | የ 4 | 47.048±0.151 | 7.815 ± 0.205 |
| 8 0 | 0 | 55.582±0.163 | 8.706± 0.215 |
| 9 88 | | 62.309±0.134 | 9.387±0.118 |
| 10 8 | 12 | 69.939±0.115 | 9.035 ± 0.205 |
| 11 | 24 | 79.578±0.213 | 9.773±0.158 |

| Table 2. Demonstrage | dmug poloogo | profile of C2 | and control gol | |
|----------------------|--------------|---------------|------------------|--|
| Table 3: Percentage | urug release | prome of G5 | and control get. | |

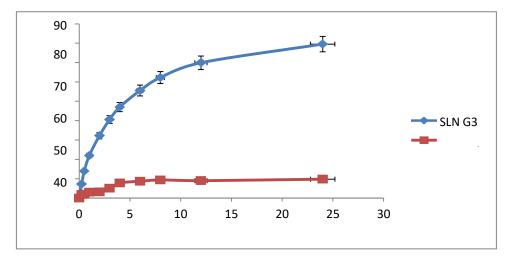
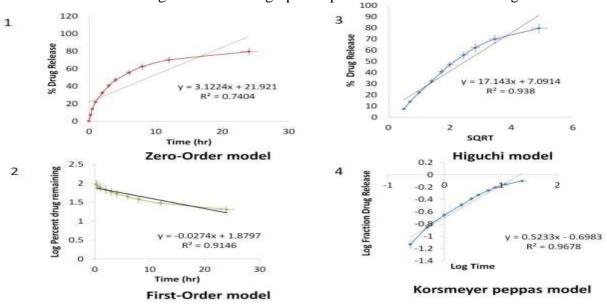


Figure 11: In-vitro drug release profile of SLN gel and control gel

Also, a medication release profile for optimal formulation applies to several kinetic models in vitro (zeroorder, first-order, Higuchi, and KrosmayerPeppas model). The collected data were statistically examined in connection to the highest correlation and rate constant to state the kinetics profiling of drug release. Except for the zero-order equation, every model contained the best-fit line. The statistics show how drugs are distributed in homogeneous matrix systems in a regulated or regular manner, and they explain why drugs spread more slowly. According to observations, SLN G3 is a considerably more effective possible topical formulation for prolonged drug delivery. As evidenced by earlier bits of information, this conclusion is virtually identical to the virtuous covenant. Figure 16 shows a graphic representation of the SLN G3 gel's kinetics order.⁸⁴





FTIR spectral analysis of SLN G3 gel

The successful FTIR study of SLN gel G3 produced spectrum data that matched those of acyclovir and stearic acid, indicating that there may be interactions between drugs and drug additives. According to spectral analysis results, the primary absorption peaks for N-H stretching are at 3331.36 cm-1, C-H stretching is at 2961.88 cm-1, C-N stretching is at 2193.49 cm-1, and C-Cl stretching is at 609.26 & 1044.56 cm-1 for



acyclovir. Although the primary absorption peaks of stearic acid have been assigned at 2932.49 cm-1 and 2863.16 cm-1 in the high-frequency area, which are attributed to asymmetric and symmetric stretching vibrations of the -CH2- band, and 1639.31 cm-1 for -COOH stretching in the low-frequency zone. Even after the consecutive creation of topical gel, spectral analysis of the improved formulation G3 demonstrates that no further drug-drug additive interactions are feasible. So, it may be argued that spectra demonstrate the SLN G3 gel's validity and purity.

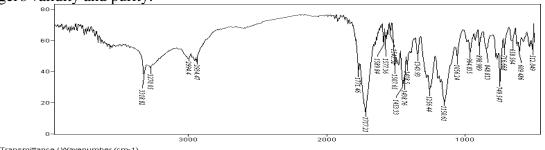


Figure13: FTIR spectra of SLN gel G3 Table 4: FTIR interpretation of SLN G3 gel.

| Characteristics Peaks | Reported (cm ⁻¹) | Observed(cm ⁻¹) |
|-----------------------|------------------------------|-----------------------------|
| N-H stretch | 3300 - 2400 | 3331.36 |
| C=C stretch | 1638 - 1648 | 1639.31 |
| CO - O – CO stretch | 1040 - 1050 | 1044. 56 |
| C – Cl stretch | 550 - 850 | 609.26 |

Scanning electron microscopy

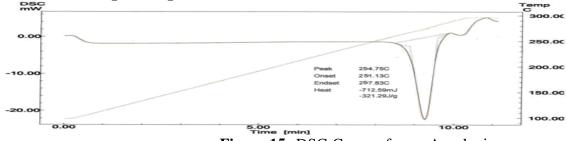
Figure 18 illustrates the increased formulation's shape after SEM analysis. The majority of vesicles are welldefined, discrete, spherical, and have extensive internal aqueous spaces. SEM examination reveals low density of nanoparticles, which may be caused by the dilution of nanosuspension prior to taking SEM images. Acyclovir-loaded SLN in gel had a spherical shape and a smooth surface, according to SEM examinations.



Figure 14: SEM analysis of SLN G3 gel

a) Differential Scanning Calorimetry (DSC)

The compatibility and interactions between drugs and polymer were checked using DSC, results obtained were shown in Figures as given.



.Figure 15: DSC Curve of pure Acyclovir

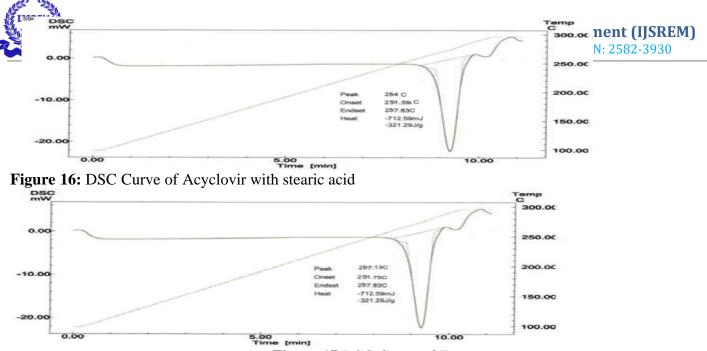


Figure 17 DSC Curve of F6

CONCLUSION

The goal of a topical drug delivery system is to deliver a therapeutic dose of medication to the appropriate location in the body, produce the intended effect, and maintain it for a period of time. To improve skin permeability and controlled drug release at the targeted site in the current study, we created solid lipid nanoparticles (SLN) that are loaded with acyclovir and added them to a topical gel of carbopol 934 that has good skin retention time. Physical and chemical characteristics of the manufactured gel were assessed in accordance with standards protocol to ensure patient compliance. There is no chemical interaction between Acyclovir and excipients, not even according to spectroscopic studies. The gel was examined under a microscope using optical and scanning electron microscopy, which revealed that SLN was distributed uniformly across the gel and that the kinetics of drug release were in good order. So, it can be said that SLN gel offers controlled drug release, and these systems can be effective as drug carriers for lipophilic pharmaceuticals, bioavailability enhancers for poorly water-soluble compounds using nanoparticles, and drug delivery systems.

References

1. Abdelbary G, Fahmy RH. Diazepam-loaded solid lipid nanoparticles: design and characterization. Aaps Pharmscitech. 2009 Mar 1;10(1):211-9.

2. Agarwal A, Agrawal H, Tiwari S, Jain S, Agrawal GP. Cationic ligand appended nanoconstructs: a prospective strategy for brain targeting. International journal of pharmaceutics. 2011 Dec 12;421(1):189-201.

3. Basha BN, Prakasam K, Goli D. Formulation and evaluation of gel containing fluconazoleantiviralagent. Int J Drug Dev Res. 2011 Oct;3(4):109-28

4. Singla V, Saini S, Joshi B, Rana AC. Emulgel: A new platform for topical drug delivery. International Journal of Pharma and Bio Sciences. 2012;3(1):485-98.

5. Bagde A, Patel K, Kutlehria S, Chowdhury N, Singh M. Formulation of topical ibuprofen solid lipid nanoparticle (SLN) gel using hot melt extrusion technique (HME) and determining its anti-inflammatory strength. Drug delivery and translational research. 2019 Mar 28:1-2

6. Bhalekar MR, Madgulkar AR, Desale PS, Marium G. Formulation of piperine solid lipid nanoparticles (SLN) for treatment of rheumatoid arthritis. Drug development and industrial pharmacy. 2017 Jun 3;43(6):1003-10.

7. Bhaskar K. Solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) enriched hydrogels for transdermal delivery of flurbiprofen: formulation, *in vitro* characterisation, pharmacokinetic and pharmacodynamic studies in animals. International Journal of Biomedical Nanoscience and Nanotechnology. 2010 Jan

1;1(2-4):109-32

8. Bhaskar K, Mohan CK, Lingam M, Mohan SJ, Venkateswarlu V, Rao YM, Bhaskar K, Anbu J, Ravichandran V. Development of SLN and NLC enriched hydrogels for transdermal delivery of nitrendipine: in vitro and *in vivo* characteristics. Drug developmentand industrial pharmacy. 2009 Jan 1;35(1):98-113. Badiee, Parisa, and Zahra Hashemizadeh. "Opportunistic invasive viralinfections: diagnosis & 9. clinical management." The Indian journal of medical research 139.2 (2014): 195. 10. Bonifaz, Alexandro, and Andrés Tirado-Sánchez. "Cutaneous disseminated and extracutaneous sporotrichosis: current status of a complex disease." Journal of Fungi 3.1 (2017): 6. Bräm S, Wolfram E, Recent Advances in Effect-directed Enzyme Assays based on Thin- layer 11. Chromatography, Phytochemical Analsis, 2017; 28(2): 74-86. Chandira RM, Pradeep PA, Bhowmik D, Chiranjib JB, Tripathi KK, Sampath Kumar KP. Design, 12. development and formulation of antiacne dermatological gel. J Chem Pharm Res. 2010;2(1):401-14. Dasgupta S, K Ghosh S, Ray S, Mazumder B. Solid lipid nanoparticles (SLNs) gels for topical 13. delivery of aceclofenac in vitro and in vivo evaluation. Current drug delivery. 2013 Dec 1;10(6):656-66. Desai, Noopur J., and Dilip G. Maheshwari, "UV spectrophotometric method for the estimation of 14. Acyclovir in marketed formulation (lotion)." Pharma Science Monitor 5.2 (2014): 48-54. 15. Deshkar SS, Bhalerao SG, Jadhav MS, Shirolkar SV. Formulation and Optimization of Topical Solid Lipid Nanoparticles Based Gel of Dapsone Using Design of Experiment. Pharmaceutical nanotechnology. 2018 Dec 1;6(4):264-75 16. Di, Li, and Edward H. Kerns. Drug-like properties: concepts, structure design and methods from ADME to toxicity optimization. Academic press, 2015. 17. Ei-Badry, M., Hassan, M. A., Ibrahim, M. A., and Elsaghir, H. (2013). Performance of poloxamer 407 as hydrophilic carrier on the binary mixtures with nimesulide. Farmacia. El-Housiny, Shaimaa, et al. "Fluconazole-loaded solid lipid nanoparticles topical gel for treatment of 18. pityriasis versicolor: formulation and clinical study." Drug delivery 25.1 (2018): 78-90. 19. El-Housiny S, Shams Eldeen MA, El-Attar YA, Salem HA, Attia D, Bendas ER, El- Nabarawi MA. Fluconazole-loaded solid lipid nanoparticles topical gel for treatment of pityriasis versicolor: formulation and clinical study. Drug delivery. 2018 Jan 1;25(1):78-90. Ekambaram, P., and Abdul Hasan Sathali, A. (2011). Formulation and evaluation of solid lipid 20. nanoparticles of ramipril. J. Young Pharm. doi:10.4103/0975-1483.83765. Ekambaram P, Sathali AA, Priyanka K. Solid lipid nanoparticles: a review. Sci Rev Chem Commun. 21. 2012 Feb;2(1):80-102 22. Felton T, Troke PF, Hope WW, Tissue penetration of antiviralagents, Clinical Microbiology Reviews, 2014; 27(1): 68-88. 23. Foldvari M. Non-invasive administration of drugs through the skin: challenges in delivery system design. Pharmaceutical science & technology today. 2000 Dec 1;3(12):417-25. Gaddam N, Aukunuru J. Systemic delivery of diclofenac sodium after topical application of gels 24. incorporated with drug-loaded solid lipid nanoparticles (SLN). Asian Journal of Pharmaceutical Research and Health Care. 2010;2(2):177-87. 25. Garse H, Jagtap P, Kadam V. Solid lipid nanoparticles based gel for topical delivery of antiviralagent. International Journal of Pharmaceutical Sciences and Research. 2015 Aug 1;6(8):3571. Gaur PK, Mishra S, Purohit S. Solid lipid nanoparticles of guggul lipid as drug carrier for 26. transdermal drug delivery. BioMed research international. 2013;2013. 27. Han F, Li S, Yin R, Shi X, Jia Q. Investigation of nanostructured lipid carriers for transdermal delivery of flurbiprofen. Drug development and industrial pharmacy. 2008 Jan1;34(4):453-8. Harde H, Agrawal AK, Katariya M, Kale D, Jain S. Development of a topical adapalene- solid lipid 28. nanoparticle loaded gel with enhanced efficacy and improved skin tolerability. RSC Advances. 2015;5(55):43917-29. Hosseini, Seyed Mostafa, et al. "Doxycycline-encapsulated solid lipid nanoparticles as promising 29. tool against Brucella melitensis enclosed in macrophage: a pharmacodynamics study on J774A. 1 cell line." Antimicrobial Resistance & Infection Control 8.1 (2019): 62. 30. https://pubchem.ncbi.nlm.nih.gov/compound/Stearic-acid

31. https://pubchem.ncbi.nlm.nih.gov/compound/Sodium-acrylate#section=FDA-Requirements.

32. https://pubchem.ncbi.nlm.nih.gov/compound/Acyclovir.

33. https://reference.medscape.com/drug/luzu-Acyclovir-999891#10

34. https://www.drugbank.ca/drugs/DB11333

35. https://www.drugbank.ca/drugs/DB08933

36. https://www.semanticscholar.org/paper/Recent-advances-in-oral-delivery-of-drugs-and-

using/b74f5b81ade6cd18889a5b063a54f8342328ab65/figure/0

37. https://www.webmd.com/drugs/2/drug-165453/Acyclovir-topical/details.

38. Hu FQ, Hong Y, Yuan H. Preparation and characterization of solid lipid nanoparticles containing peptide. International journal of pharmaceutics. 2004 Apr 1;273(1-2):29-35.

39. Ige, Pradum Pundlikrao, Rohan K. Baria, and Surendra G. Gattani. "Fabrication of fenofibrate nanocrystals by probe sonication method for enhancement of dissolution rate and oral bioavailability." *Colloids and Surfaces B: Biointerfaces* 108 (2013): 366-373.

40. Jain A, Garg NK, Jain A, Kesharwani P, Jain AK, Nirbhavane P, Tyagi RK. A synergistic approach of adapalene-loaded nanostructured lipid carriers, and vitamin C co- administration for treating acne. Drug development and industrial pharmacy. 2016 Jun 2;42(6):897-905.

41. Jana, Sougata, et al. "Carbopol gel containing chitosan-egg albumin nanoparticles for transdermal aceclofenac delivery." *Colloids and surfaces B: Biointerfaces* 114 (2014): 36-44.

42. Jenning V, Gysler A, Schäfer-Korting M, Gohla SH. Vitamin A loaded solid lipid nanoparticles for topical use: occlusive properties and drug targeting to the upper skin. European journal of pharmaceutics and biopharmaceutics. 2000 May 2;49(3):211-8.

43. Kang KW, Chun MK, Kim O, Subedi RK, Ahn SG, Yoon JH, Choi HK. Doxorubicin- loaded solid lipid nanoparticles to overcome multidrug resistance in cancer therapy. Nanomedicine: Nanotechnology, Biology and Medicine. 2010 Apr 1;6(2):210-3.

44. Kansagra H, Mallick S. Microemulsion-based antiviralgel of Acyclovir for dermatophyte infections: formulation, characterization and efficacy studies. Journal of Pharmaceutical Investigation. 2016 Feb 1;46(1):21-8.

45. Kaur M, Singh K, Jain SK. Acyclovir vesicular based gel formulations for its enhanced topical delivery. Journal of liposome research. 2019 Oct 17(just-accepted):1-43.

46. Kaur J, Kaur J, Jaiswal S, Gupta GD. Recent advances in topical drug delivery system. Pharmaceutical Research. 2016;6(07).

47. Kesharwani R, Sachan A, Singh S, Patel D. Formulation and evaluation of solid lipid nanoparticle (SLN) based topical gel of etoricoxib. Journal of Applied Pharmaceutical Science. 2016 Oct;6(10):124-31.

48. Khanna, D., and S. Bharti. "Acyclovir for the treatment of viralinfections: An evidence-based review. Core Evid. 2014; 9: 113–24."

49. Koga, Hiroyasu, et al. "Short-term therapy with Acyclovir, a novel topical antiviralimidazole, in guinea pig models of tinea corporis and tinea pedis." *Antimicrobial agents and chemotherapy* 56.6 (2012): 3138-3143.

50. Krishnatreyya H, Dey S, Pal P, Das PJ, Sharma VK, Mazumder B. Piroxicam Loaded Solid Lipid Nanoparticles (SLNs): Potential for Topical Delivery. INDIAN JOURNAL OF PHARMACEUTICAL EDUCATION AND RESEARCH. 2019 Apr 1;53(2):S82-92.

51. Kumar, Manish, et al. "Preparation of Acyclovir nanocrystals loaded hydrogel for improvement of dissolution and antiviralactivity." *Heliyon* 5.5 (2019): e01688.

52. Kumar M, Shanthi N, Mahato AK, Soni S, Rajnikanth PS. Preparation of Acyclovir nanocrystals loaded hydrogel for improvement of dissolution and antiviralactivity. Heliyon. 2019 May 1;5(5):e01688.

53. Lakshmi CV, Bengalorkar GM, Kumar VS. Clinical efficacy of topical terbinafine versus topical Acyclovir in treatment of tinea corporis/tinea cruris patients. Journal of Pharmaceutical Research International. 2013 Aug 24:1001-14.

54. Liu W, Hu M, Liu W, Xue C, Xu H, Yang X. Investigation of the carbopol gel of solid lipid nanoparticles for the transdermal iontophoretic delivery of triamcinolone acetonide acetate. International journal of pharmaceutics. 2008 Nov 19;364(1):135-41.

55. Madan, Jyotsana R., Priyanka A. Khude, and Kamal Dua. "Development and evaluation of solid lipid nanoparticles of mometasone furoate for topical delivery." *International journal of pharmaceutical investigation* 4.2



(2014): 60.

59.

56. Makri, Nikoleta, et al. "First case report of cutaneous sporotrichosis (Sporothrix species) in a cat in the UK." *Journal of Feline Medicine and Surgery Open Reports* 6.1 (2020): 2055116920906001.

57. Mukherjee, S., S. Ray, and R. S. Thakur. "Solid lipid nanoparticles: a modern formulation approach in drug delivery system." *Indian journal of pharmaceutical sciences* 71.4 (2009):349.

58. Muller, R. "Solid lipid nanoparticles (SLN) for controlled drug delivery –a review of the state of the art." *Eur. J. Pharm. Biopharm* 50.1 (2000): 161-177.

Mali AD. An updated review on transdermal drug delivery systems. Skin. 2015;8(9).

60. Naseri N, Valizadeh H, Zakeri-Milani P, Solid lipid nanoparticles and nanostructured lipid carriers: Structure preparation and application, Advanced Pharmaceutical Bulletin, 2015; 5(3): 305–313.

61. Ng KW, Lau WM. Skin deep: the basics of human skin structure and drug penetration. In Percutaneous penetration enhancers chemical methods in penetration enhancement 2015 (pp. 3-11). Springer, Berlin, Heidelberg.

62. Paliwal, R., Rai, S., Vaidya, B., Khatri, K., Goyal, A. K., Mishra, N., et al. (2009). Effect of lipid core material on characteristics of solid lipid nanoparticles designed for oral lymphatic delivery. Nanomedicine Nanotechnology, Biol. Med. doi:10.1016/j

63. Pandurangan, Dinesh Kumar, et al. "Formulation and evaluation of voriconazole ophthalmic solid lipid nanoparticles in situ gel." *International journal of pharmaceutical investigation* 6.1 (2016): 56.

64. Pawar PM, Solanki KP, Mandali VA. Recent Advancements in Transdermal Drug Delivery System. International Journal of Pharmaceutical and Clinical Research.2018;10(3):65-73.

65. Pople PV, Singh KK. Development and evaluation of topical formulation containing solid lipid nanoparticles of vitamin A. Aaps Pharmscitech. 2006 Dec 1;7(4):E63-9

66. Pudney, Paul DA, Kevin J. Mutch, and Shiping Zhu. "Characterising the phase behaviour of stearic acid and its triethanolamine soap and acid–soap by infrared spectroscopy." *Physical Chemistry Chemical Physics* 11.25 (2009): 5010-5018.

67. Radomska-Soukharev A. Stability of lipid excipients in solid lipid nanoparticles. Advanced drug delivery reviews. 2007 Jul 10;59(6):411-8.

68.Ramteke KH, Joshi SA, Dhole SN. Solid lipid nanoparticle: A review. IOSR J. Pharm. 2012;2(6):34-44.

69. Raghava Srivalli KM, Mishra B, Drug nanocrystals: A way toward scale-up, Saudi Pharmaceutical Journal, 2016; 24(4): 386–404.

70. Raza K, Singh B, Singal P, Wadhwa S, Katare OP. Systematically optimized biocompatible isotretinoin-loaded solid lipid nanoparticles (SLNs) for topical treatment of acne. Colloids and Surfaces B: Biointerfaces. 2013 May 1;105:67-74.

71. Reddigari JR, Ramesh Y, Kothapalli CB. Formulation and evaluation of in-situ gels enriched with Tropicamide loaded solid lipid nanoparticles. International Journal of Research in Pharmaceutical sciences. 2018 Jan 10;9(1):216-25.

72. Sanna V, Gavini E, Cossu M, Rassu G, Giunchedi P. Solid lipid nanoparticles (SLN) as carriers for the topical delivery of econazole nitrate: in-vitro characterization, ex-vivo and in-vivo studies. Journal of pharmacy and pharmacology. 2007 Aug;59(8):1057-64.

73. Sahni, Kanika, Sanjay Singh, and Sunil Dogra. "Newer topical treatments in skin and nail dermatophyte infections." *Indian dermatology online journal* 9.3 (2018): 149.

74. SARANGI MK, PADHI S. Solid lipid nanoparticles–a review. drugs. 2016;5:7.

75. Shah, M., et al. "Solid lipid nanoparticles of a water soluble drug, ciprofloxacin hydrochloride." *Indian journal of pharmaceutical sciences* 74.5 (2012): 434.

76. Shah KA, Date AA, Joshi MD, Patravale VB. Solid lipid nanoparticles (SLN) of tretinoin: potential in topical delivery. International journal of pharmaceutics. 2007 Dec 10;345(1-2):163-71.

77. Singh AP, Saraf SK, Saraf SA. SLN approach for nose-to-brain delivery of alprazolam. Drug delivery and translational research. 2012 Dec 1;2(6):498-507.

78. Singla V, Saini S, Joshi B, Rana AC. Emulgel: A new platform for topical drug delivery. International Journal of Pharma and Bio Sciences. 2012;3(1):485-98.



79. Srivalli, Kale Mohana Raghava, and Brahmeshwar Mishra. "Drug nanocrystals: A way toward scaleup." *Saudi Pharmaceutical Journal* 24.4 (2016): 386-404.

80. Stella, Barbara, et al. "Development and characterization of solid lipid nanoparticles loaded with a highly active doxorubicin derivative." *Nanomaterials* 8.2 (2018): 110.

81. Tang, Jingling, et al. "Solid lipid nanoparticles with TPGS and Brij 78: a co-delivery vehicle of curcumin and piperine for reversing P-glycoprotein-mediated multidrug resistance in vitro." *Oncology Letters* 13.1 (2017): 389-395.

82. Tanwar H, Sachdeva R. Transdermal drug delivery system: A review. Int. J. Pharm. Sci. Res. 2016 Jun 1;7:2274-90.

83. Tian, Huaixiang, et al. "Preparation and characterization of citral-loaded solid lipid nanoparticles." *Food chemistry* 248 (2018): 78-85.

84. Tiyaboonchai W, Tungpradit W, Plianbangchang P. Formulation and characterization of curcuminoids loaded solid lipid nanoparticles. International Journal of Pharmaceutics. 2007 Jun 7;337(1-2):299-306.

85. Tyagi RK, Chandra A, Singh D, Rahman MA. Transdermal drug delivery system (TDDS): an overview. International journal of pharmaceutical sciences and research. 2011 Jun 1;2(6):1379-88.

86. Uprit, Shubham, et al. "Preparation and characterization of minoxidil loaded nanostructured lipid carrier gel for effective treatment of alopecia." *Saudi Pharmaceutical Journal* 21.4 (2013): 379-385.

87. Vidlářová, Lucie, et al. "Nanocrystals for dermal penetration enhancement–effect of concentration and underlying mechanisms using curcumin as model." *European Journal of Pharmaceutics and Biopharmaceutics* 104 (2016): 216-225.

88. Yusuf, Mohammad, et al. "Plausible antioxidant biomechanics and anticonvulsant pharmacological activity of brain-targeted β-carotene nanoparticles." *International journal of nanomedicine* 7 (2012): 4311.